


# Introductory Guide to Light Microscopy

## - Biomedical Confocal Microscopy

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$$E = hv$$


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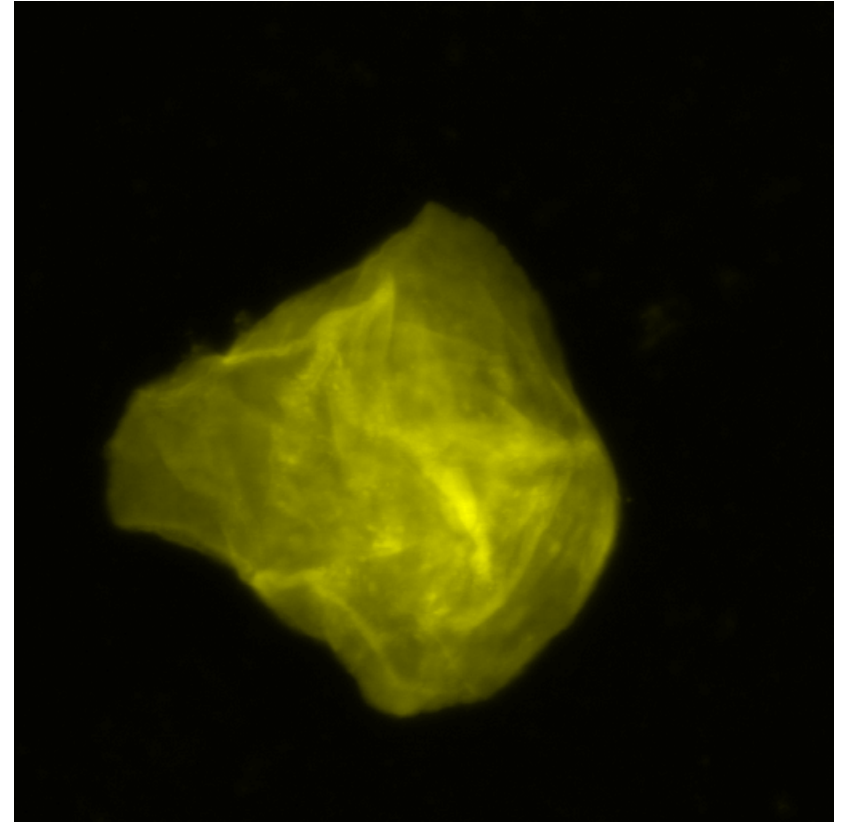
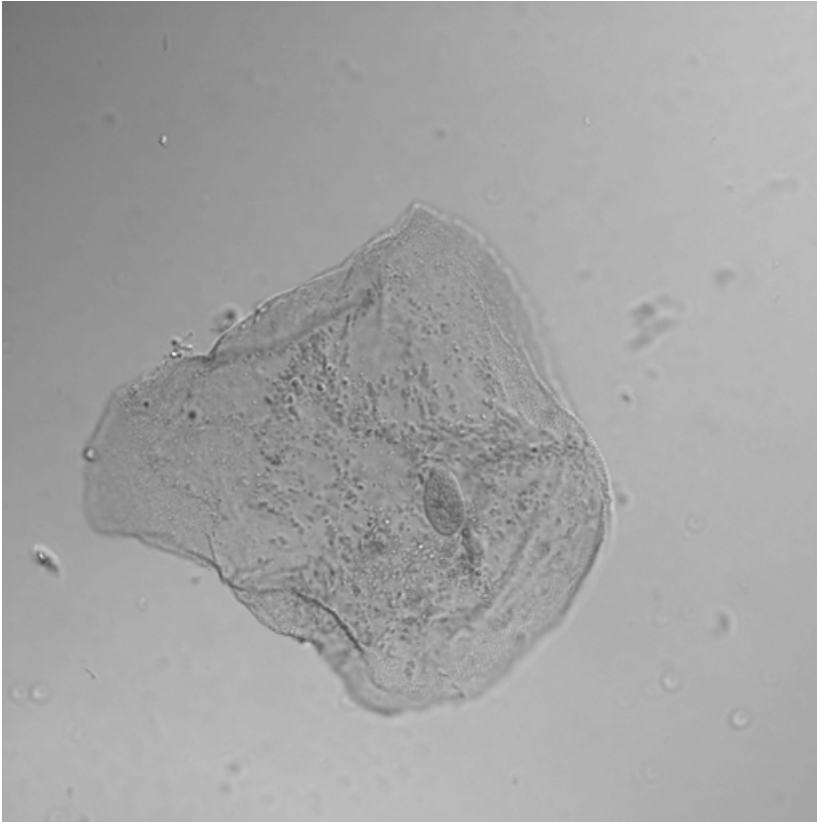
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# Confocal Microscopy

- Limitation of wide field microscopy
- Origins of Confocal Microscopy
- Confocal Principal
- Confocal Laser Scanning
- Resolution
- X-Z axis scanning
- Visualization



# Limitation of wide field microscopy



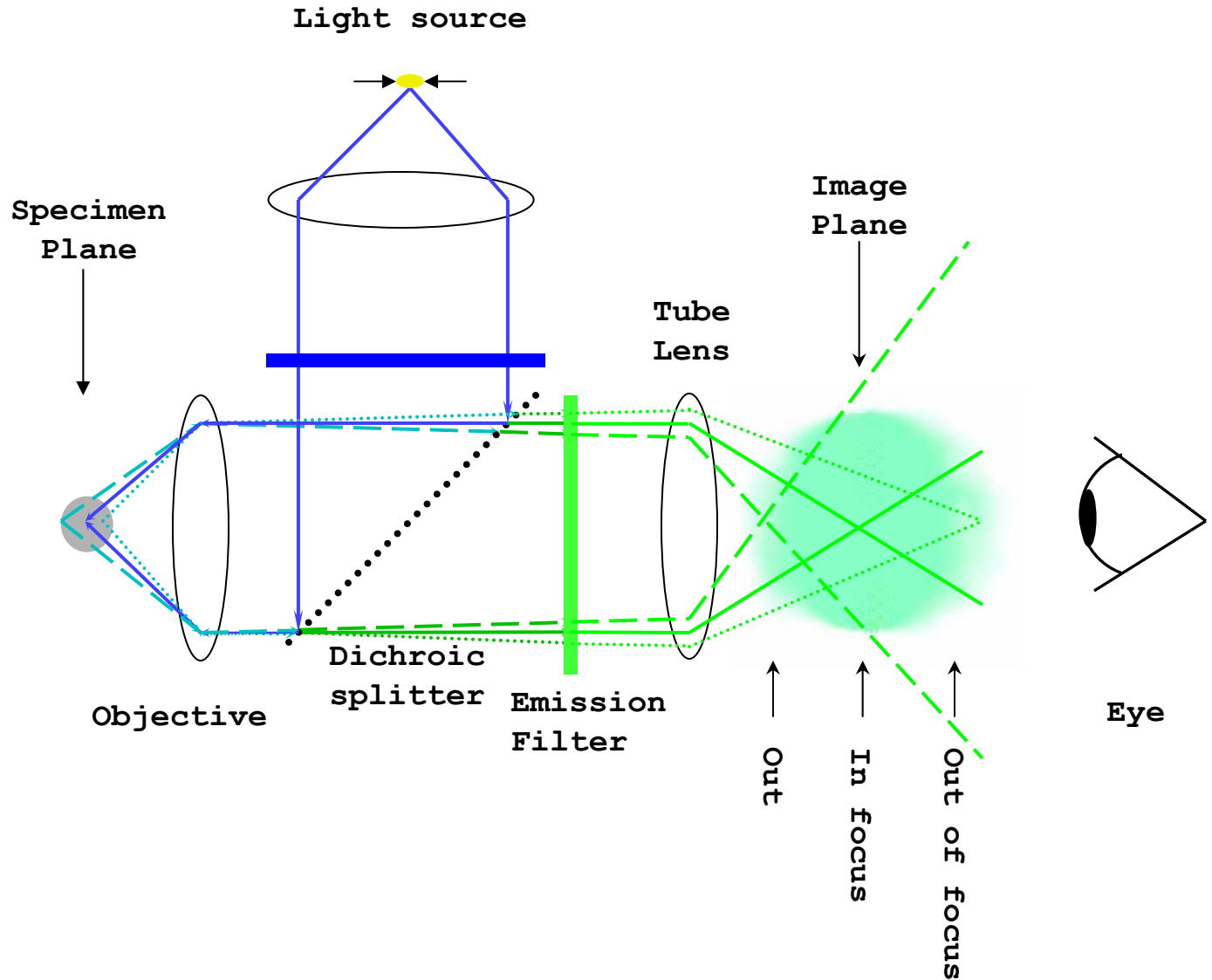
Live Buccal Epithelial cells

(DIC) Transmitted

Fluorescence

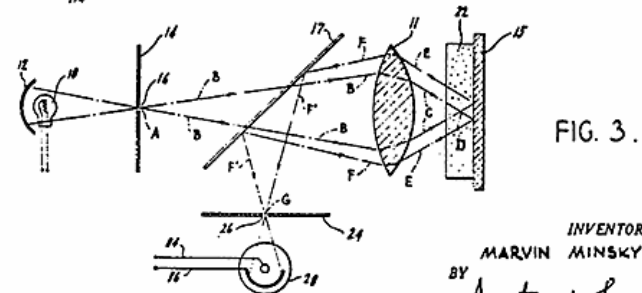
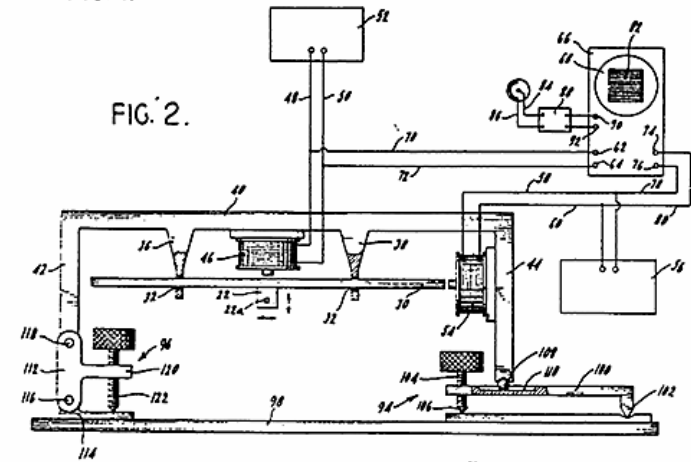
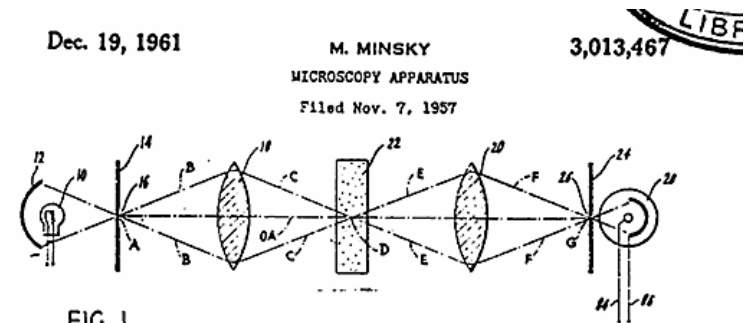
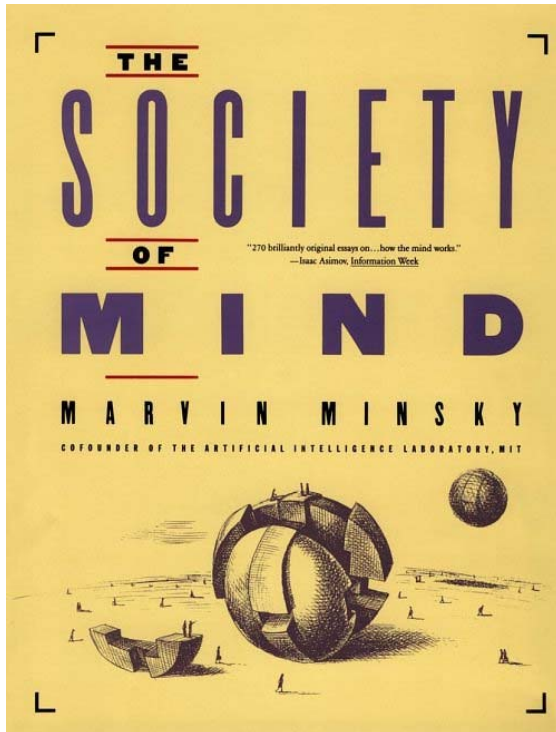
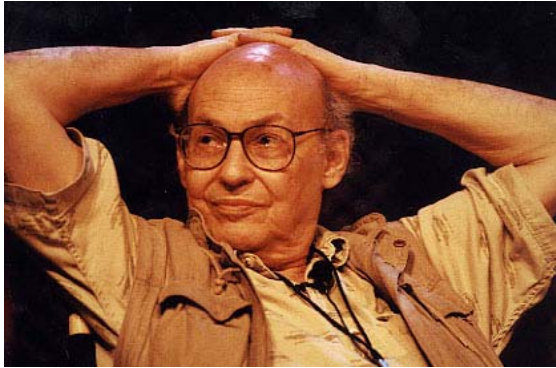
FM 1-43 membrane dye

# Limitation of wide field microscopy



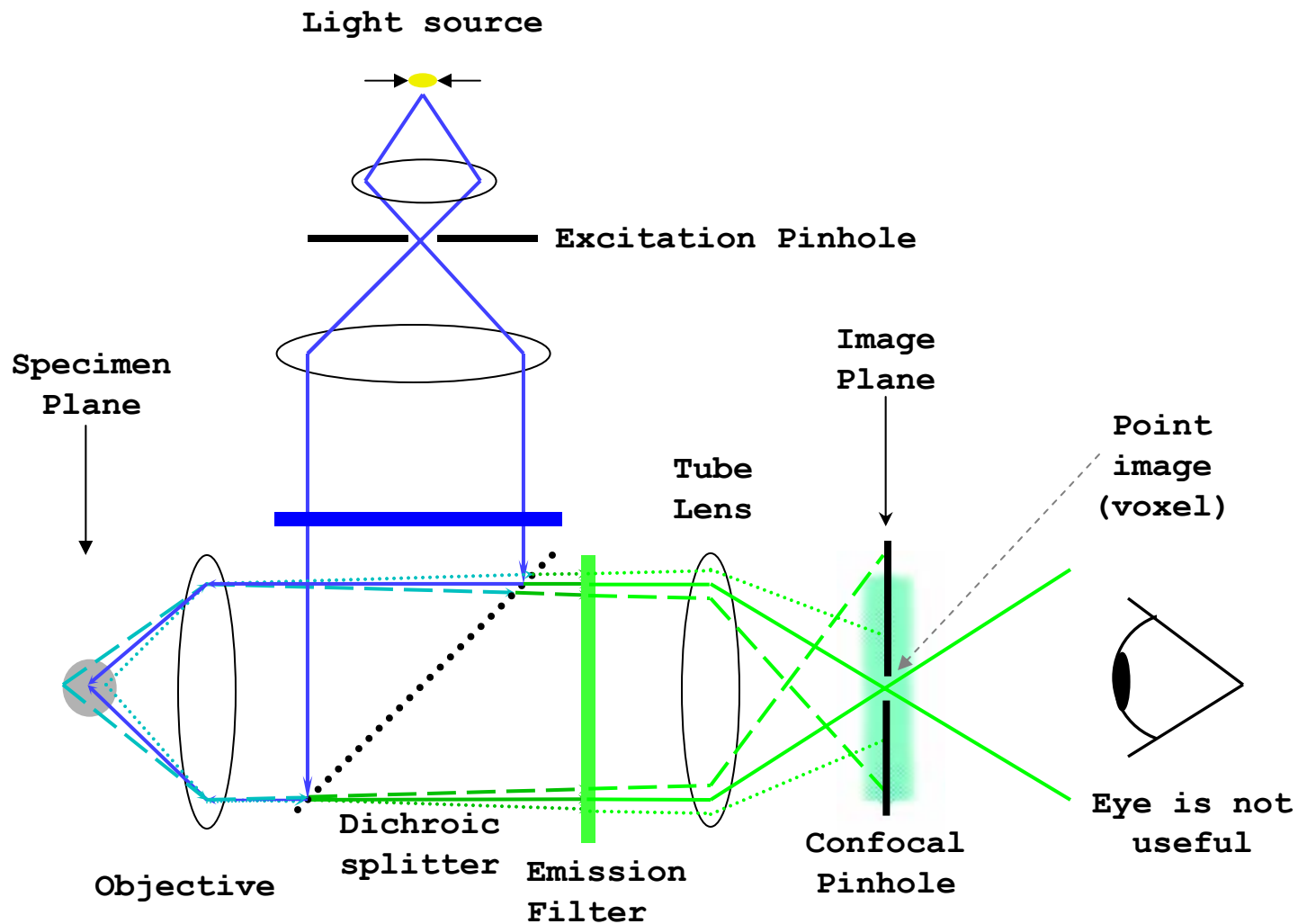
# Origins – Marvin Minsky

1957 Confocal Patent focal Scanning Microscope: U.S. Patent 3013467

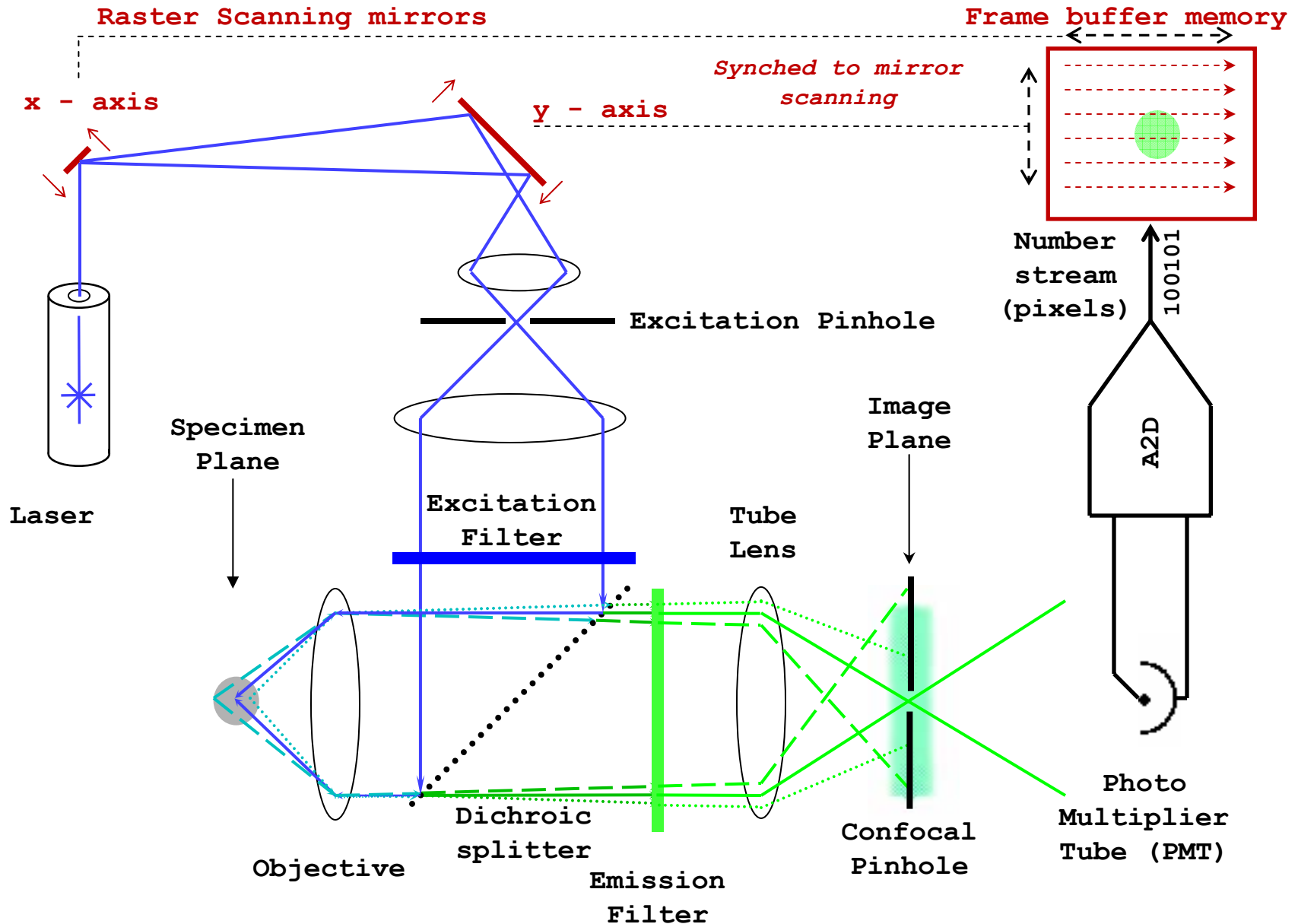


INVENTOR  
MARVIN MINSKY  
BY  
*Ametor & Levy*

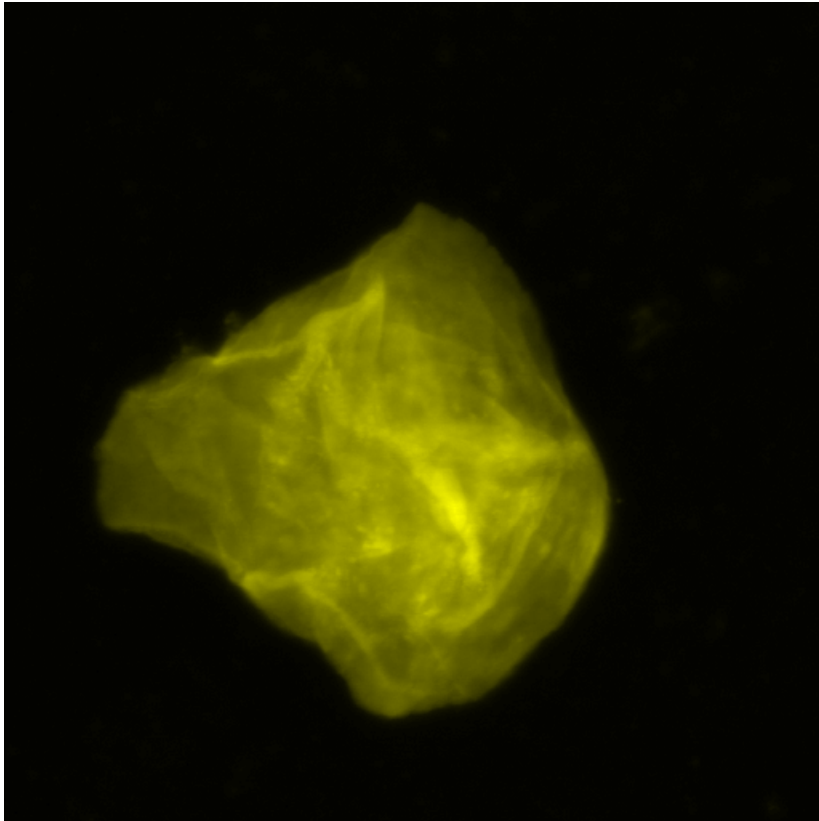
# Confocal Principal – the confocal pinhole



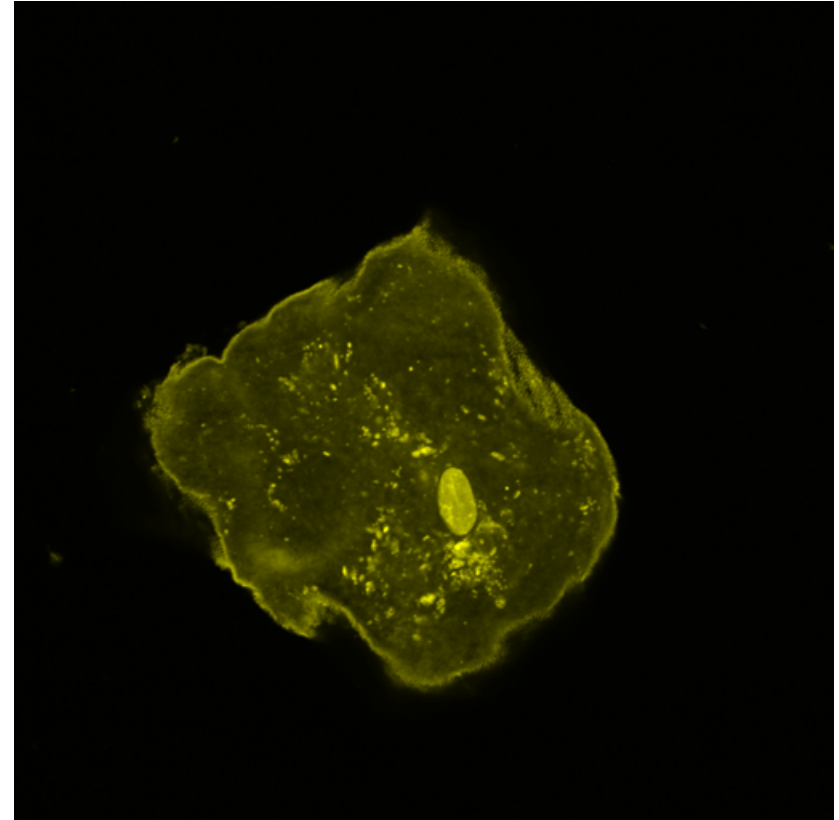
# Confocal Laser Scanning Microscope



# Confocal Principal – Live Buccal Epithelial cells



Widefield



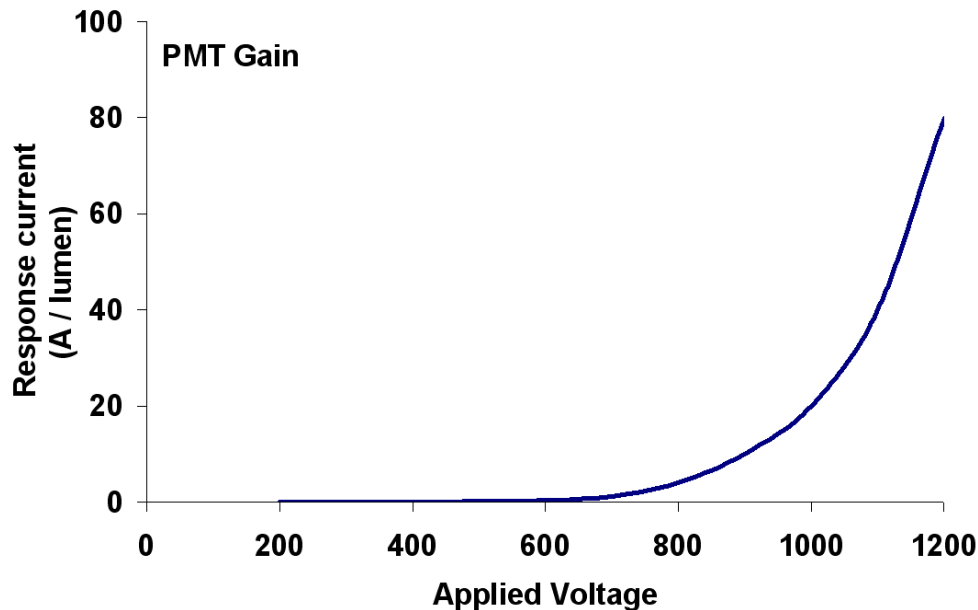
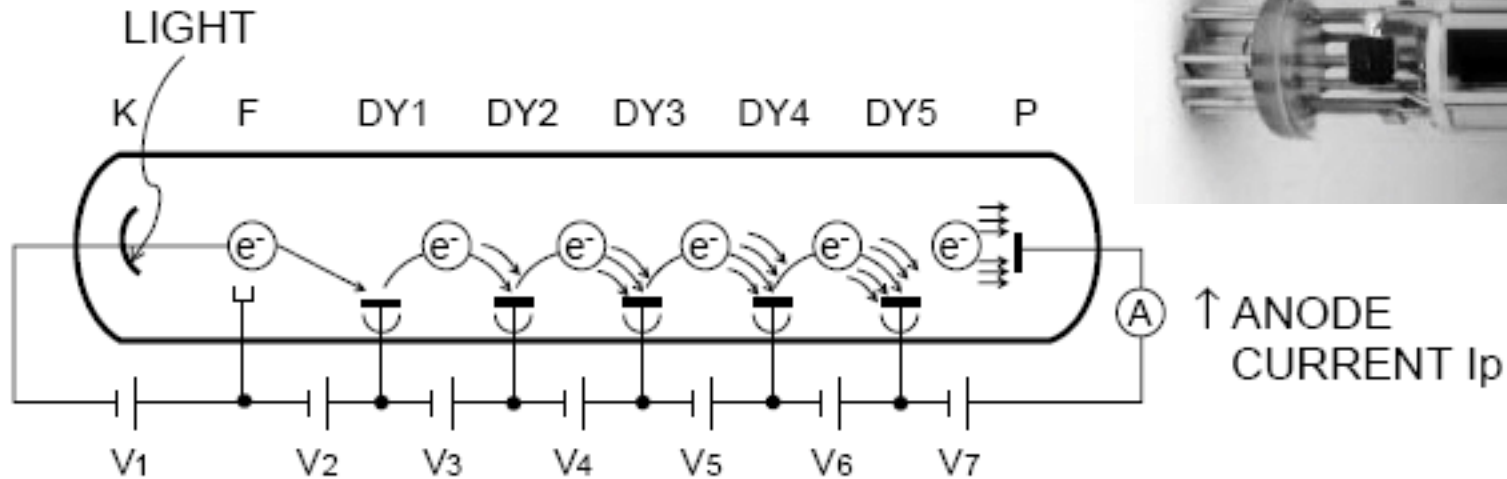
Confocal

FM 1-43 membrane dye



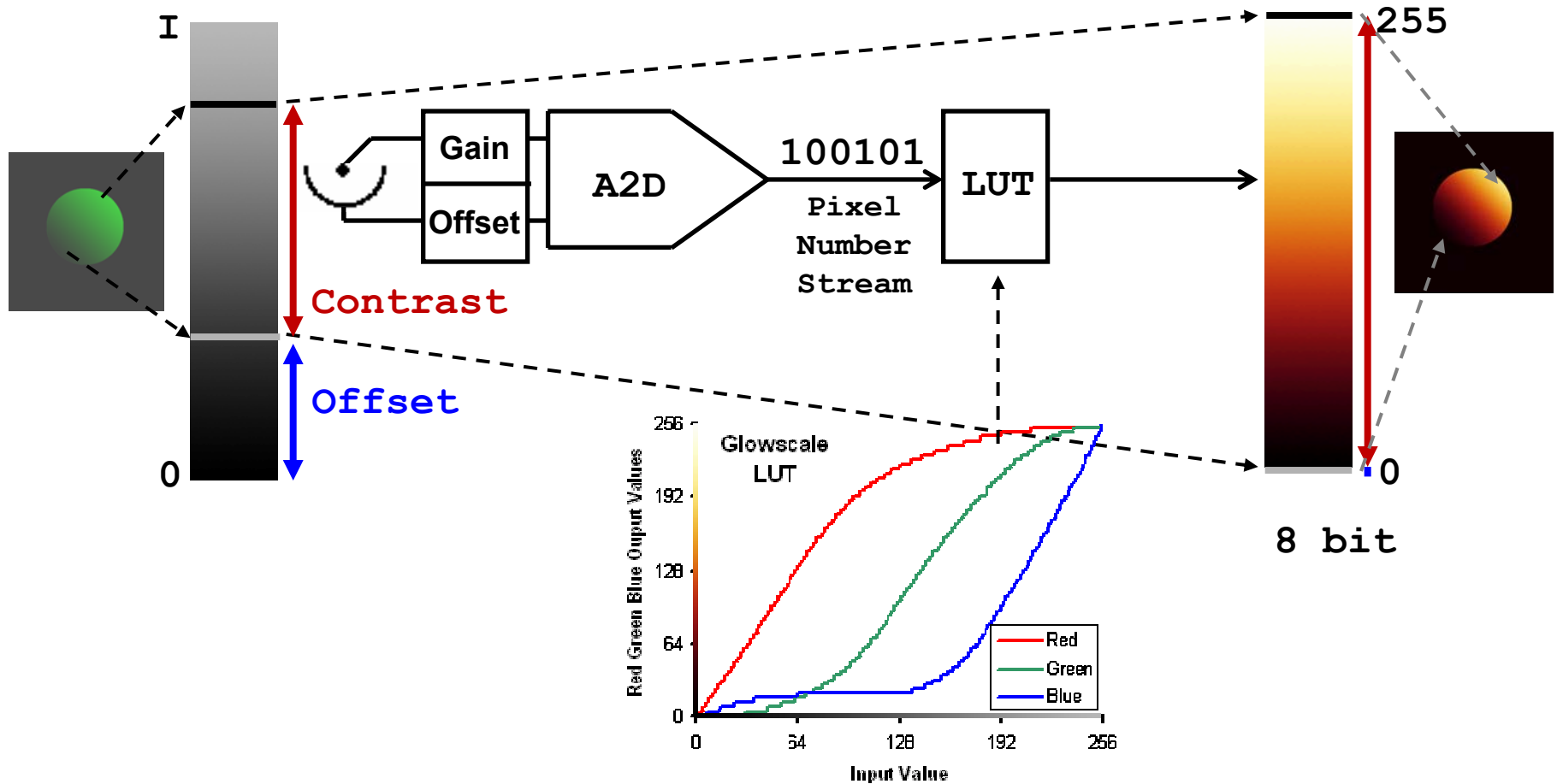
# Confocal Laser Scanning Microscope – PMT

Photo Multiplier Tube (PMT)



- Quantum yield  $\leq 0.3$
- Gain can be very large  $>10^7$
- Gain is exponential function of applied voltage
- Noise increases disproportionately at high gain
- Significant dark current

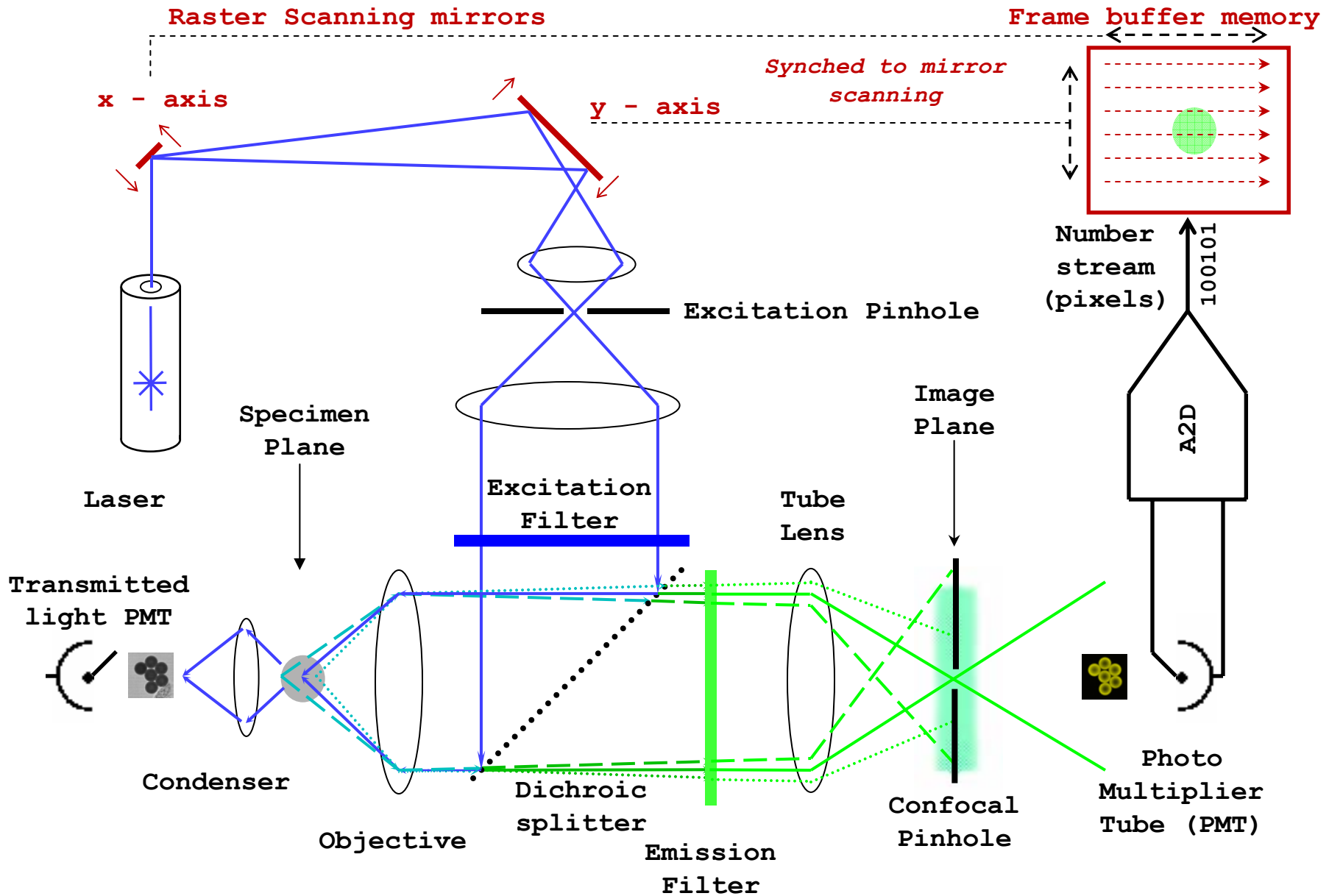
# Confocal Laser Scanning Microscope– Analogue to Digital Converter & Frame Buffer Display



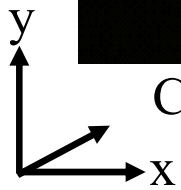
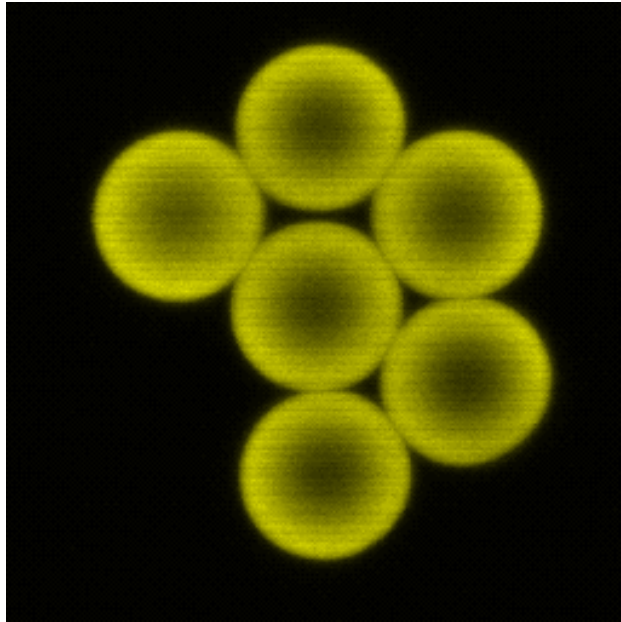
A2D = Analogue to Digital Converter

LUT = Look Up Table

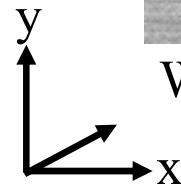
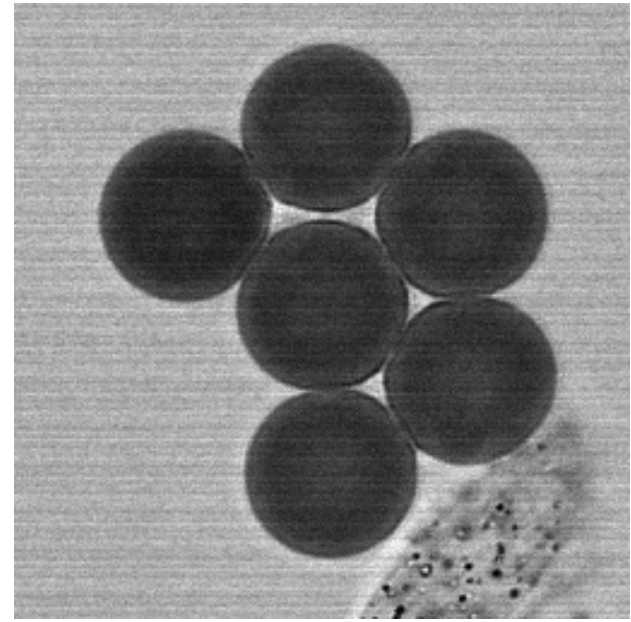
# Confocal Laser Scanning Microscope



# 10 um beads – xy view

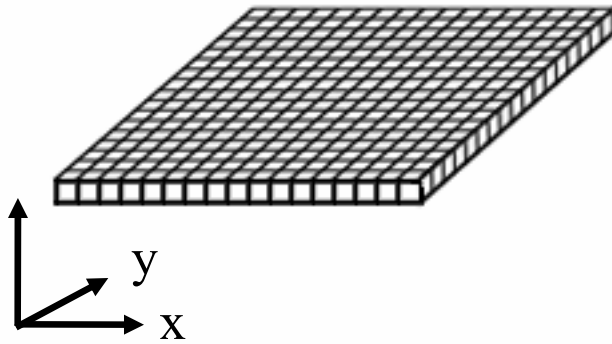


Confocal fluorescence

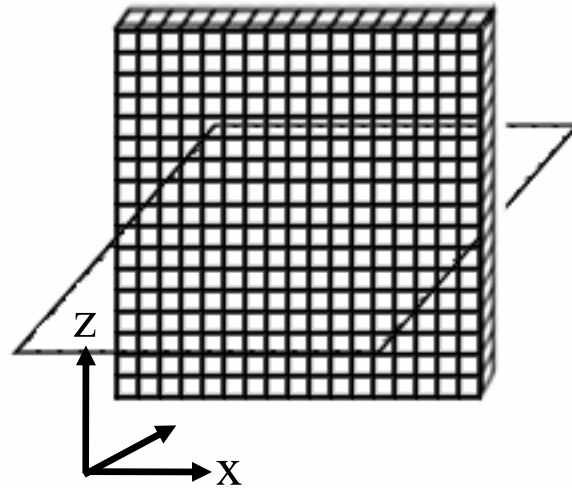


Wide field transmitted light

# Scanning modes



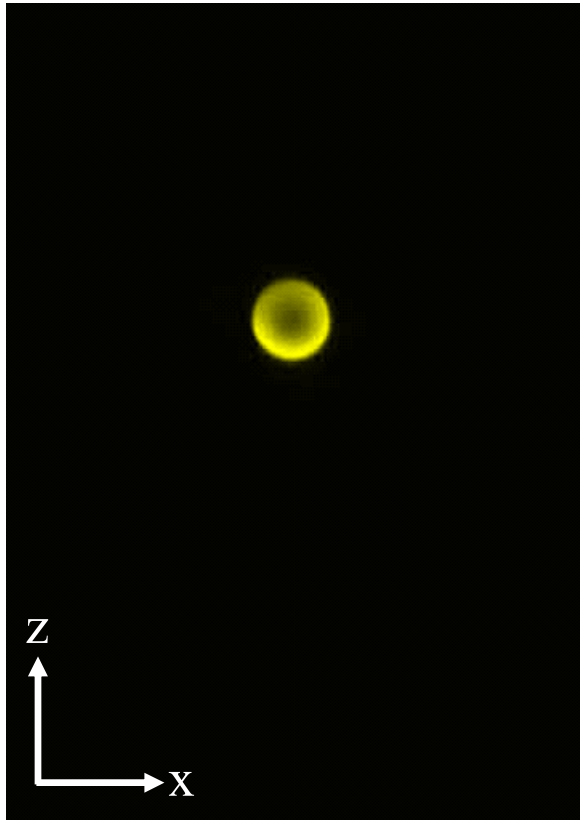
x - y scanning  
(conventional slice)



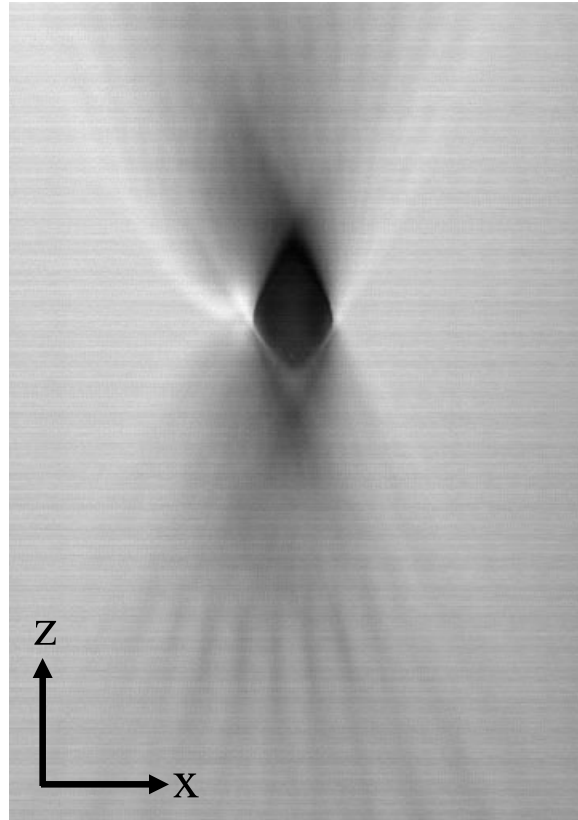
x - z scanning  
(vertical slice)

↑  
↓  
Stage  
vibrated

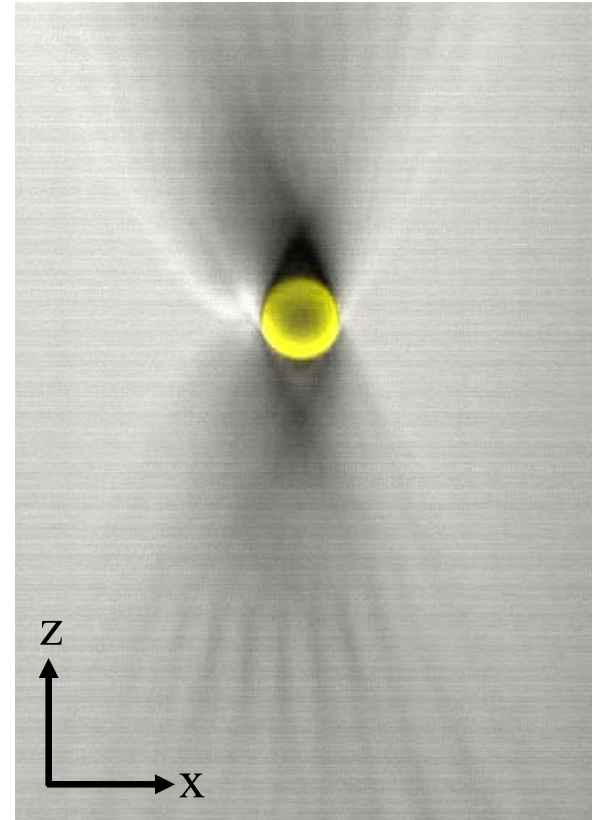
# 10 um bead – xz side view



Confocal fluorescence

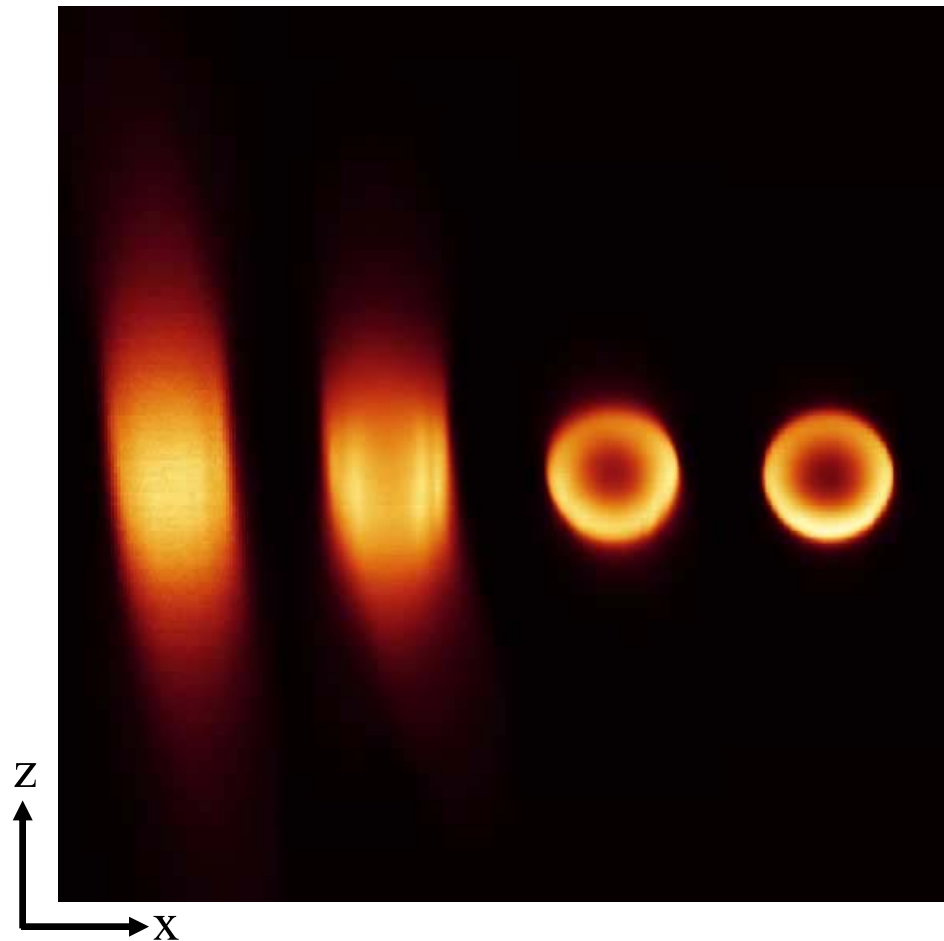


Wide field transmitted light



Overlay

# 10 um bead – confocal xz view



10x

20x

40x

100x

objective

0.3

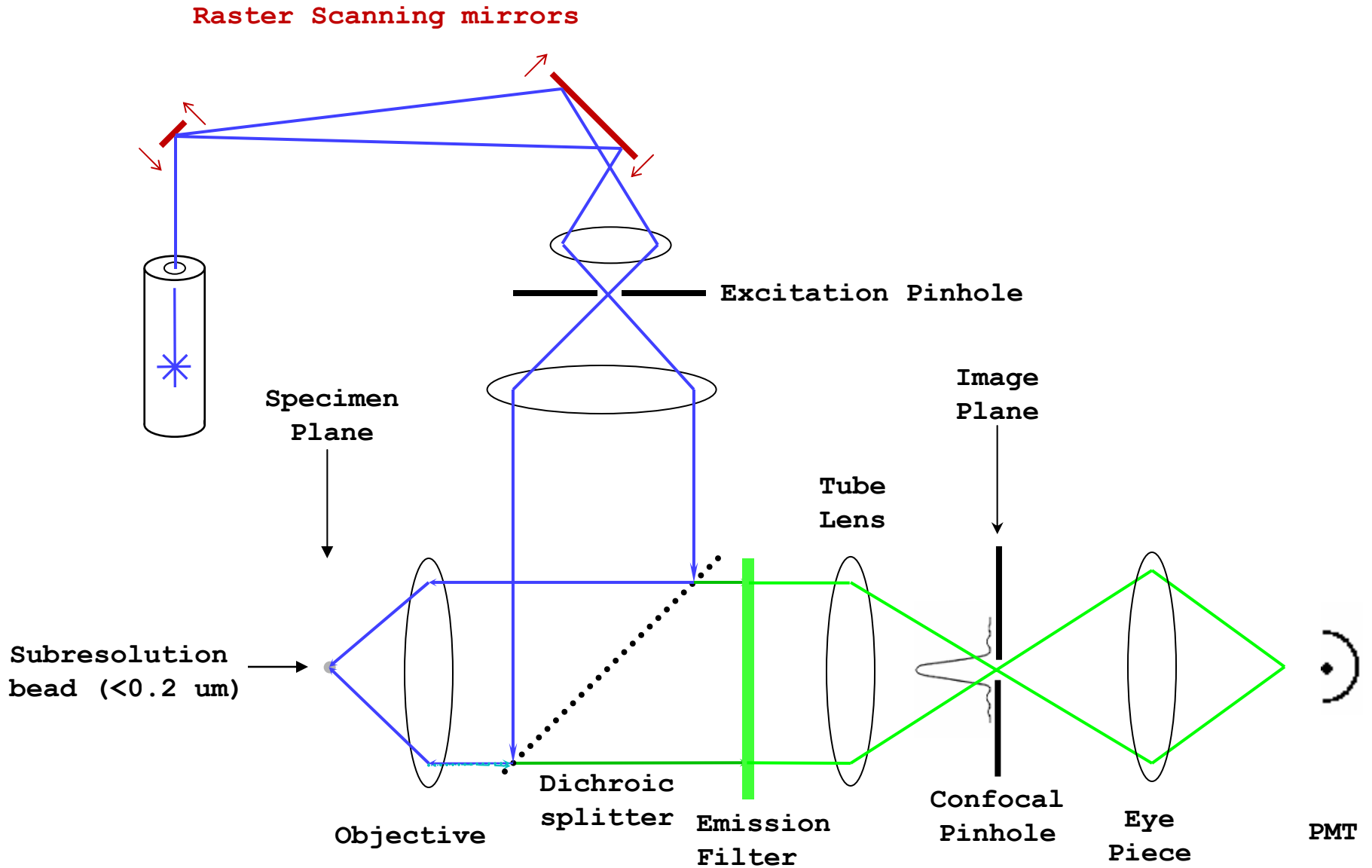
0.7

1.25

1.4

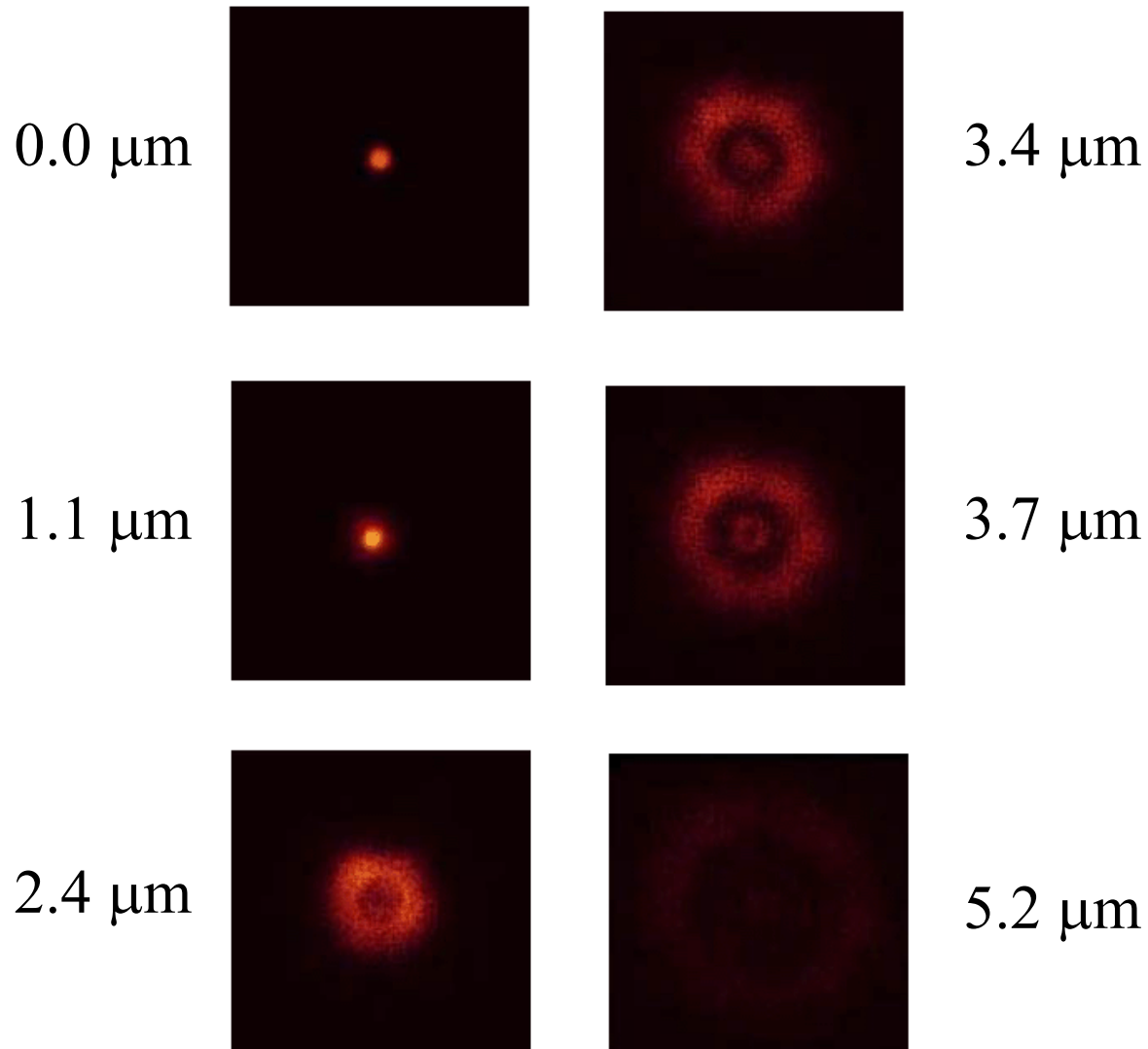
numerical aperture

# Airy Disk



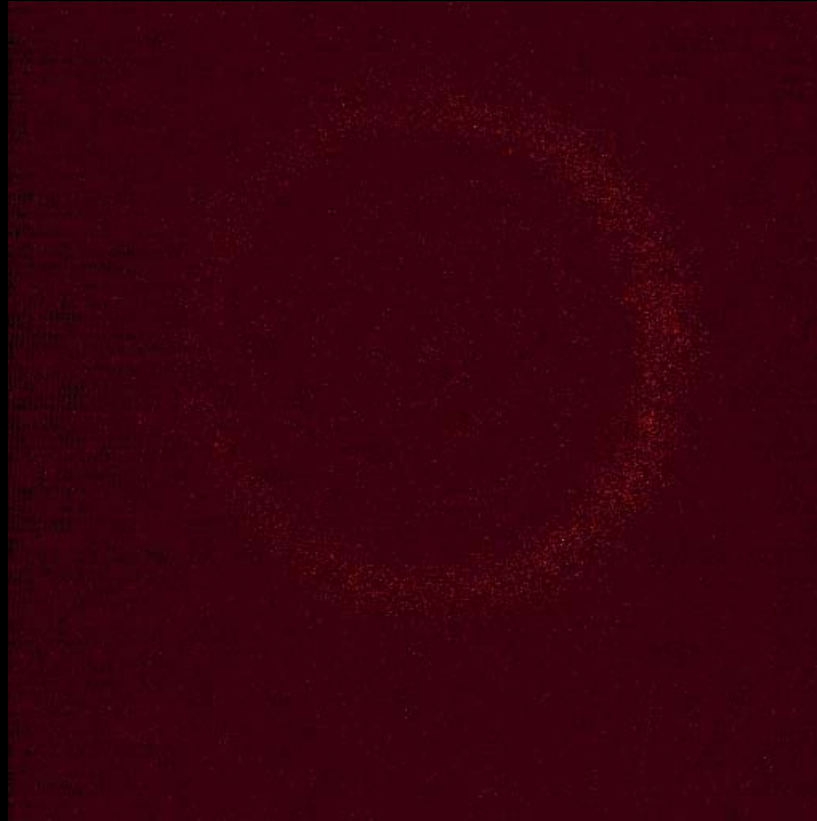


# Airy Disks



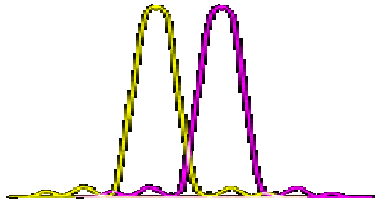
0.5  $\mu\text{m}$  bead Plan Apo 100x 1.4 NA oil

# Airy Disk



0.5 um bead Plan Apo 20x 0.7 NA oil

# Numerical Aperture (NA) - resolution



Rayleigh Criterion: 2 points are said to be resolved when their separation causes the center of the Airy pattern of one point to fall on the 1<sup>st</sup> minimum of the other Airy disk

$$NA = n \cdot \sin(a), \quad a = \text{half cone angle}$$

$n$  = refractive index of medium  
 $\lambda$  = wavelength

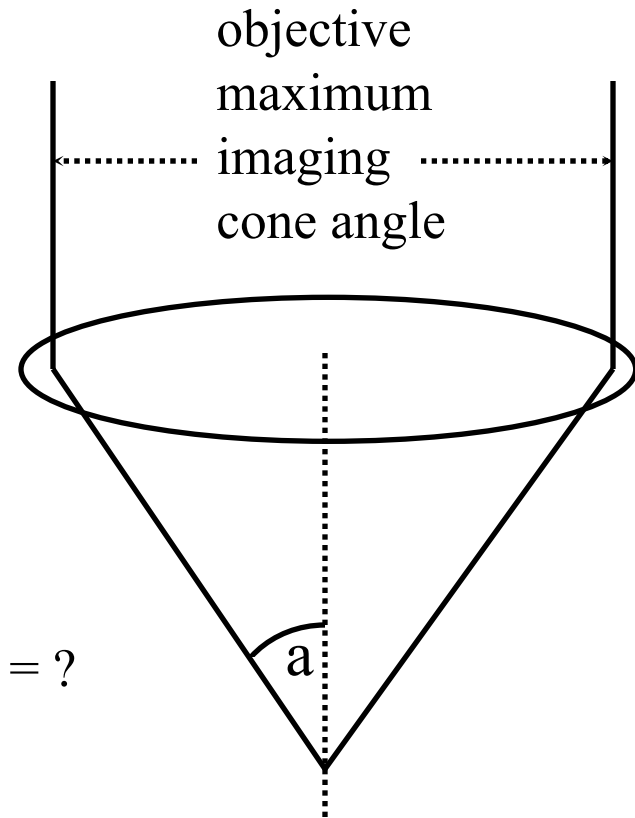
Wide Field (fluorescence)  $r_{\text{airy}} = 0.61 \lambda / NA$   
 $= 0.22 \mu\text{m}$

Confocal  $r_{\text{airy}} = 0.61 \lambda / NA / \sqrt{2}$   
 $= 0.15 \mu\text{m}$

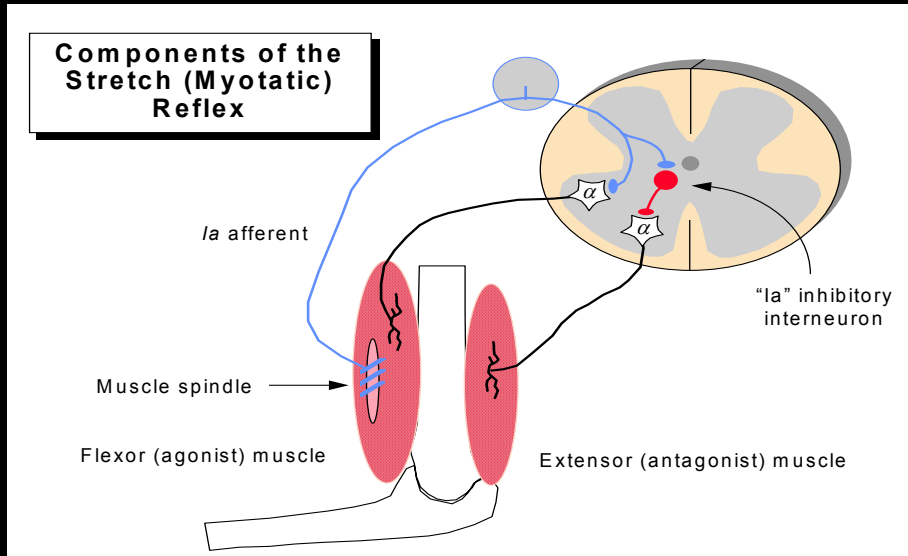
$$r_{\text{airy.axial}} = ?$$

$$r_{\text{axial-fluor}} = 1.77 \lambda / NA^2$$

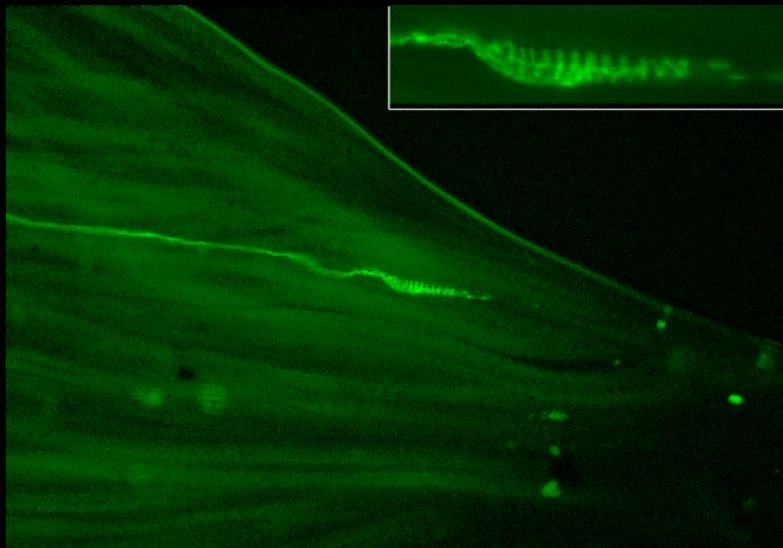
$= 0.4 \mu\text{m}$  (for green emission)



# Visualization - Example Muscle Spindle



Transmitted light 16x widefield



Fluorescence 4x widefield transgenic mouse Thy1-YFG

Wide field – non confocal

Thin slice and z-series

Gallery

Serial slices

Z-axis sampling

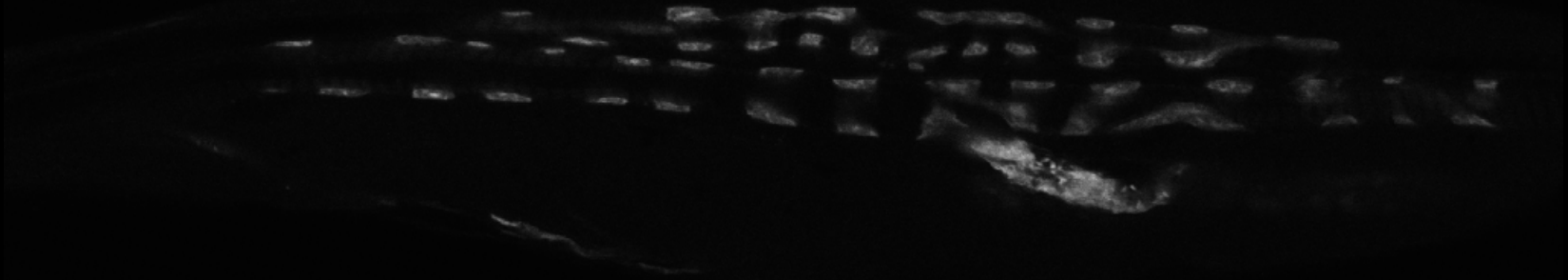
Orthogonal view

Average & Maximum projections

Volume render

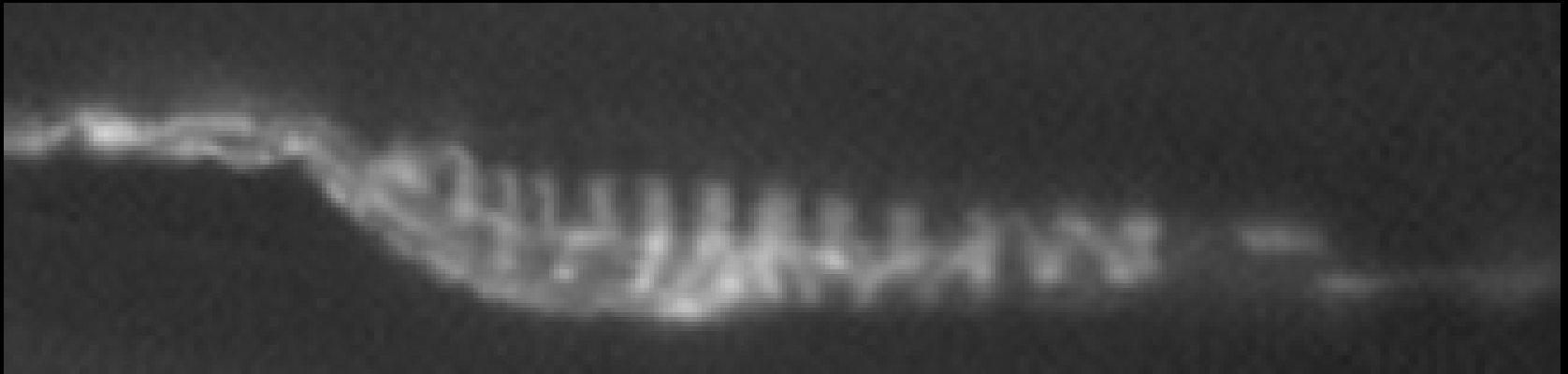
Multiple channels (dyes / fluorophores)

# Visualization - Live Muscle Spindle (Thy1-YFP)



single confocal slice

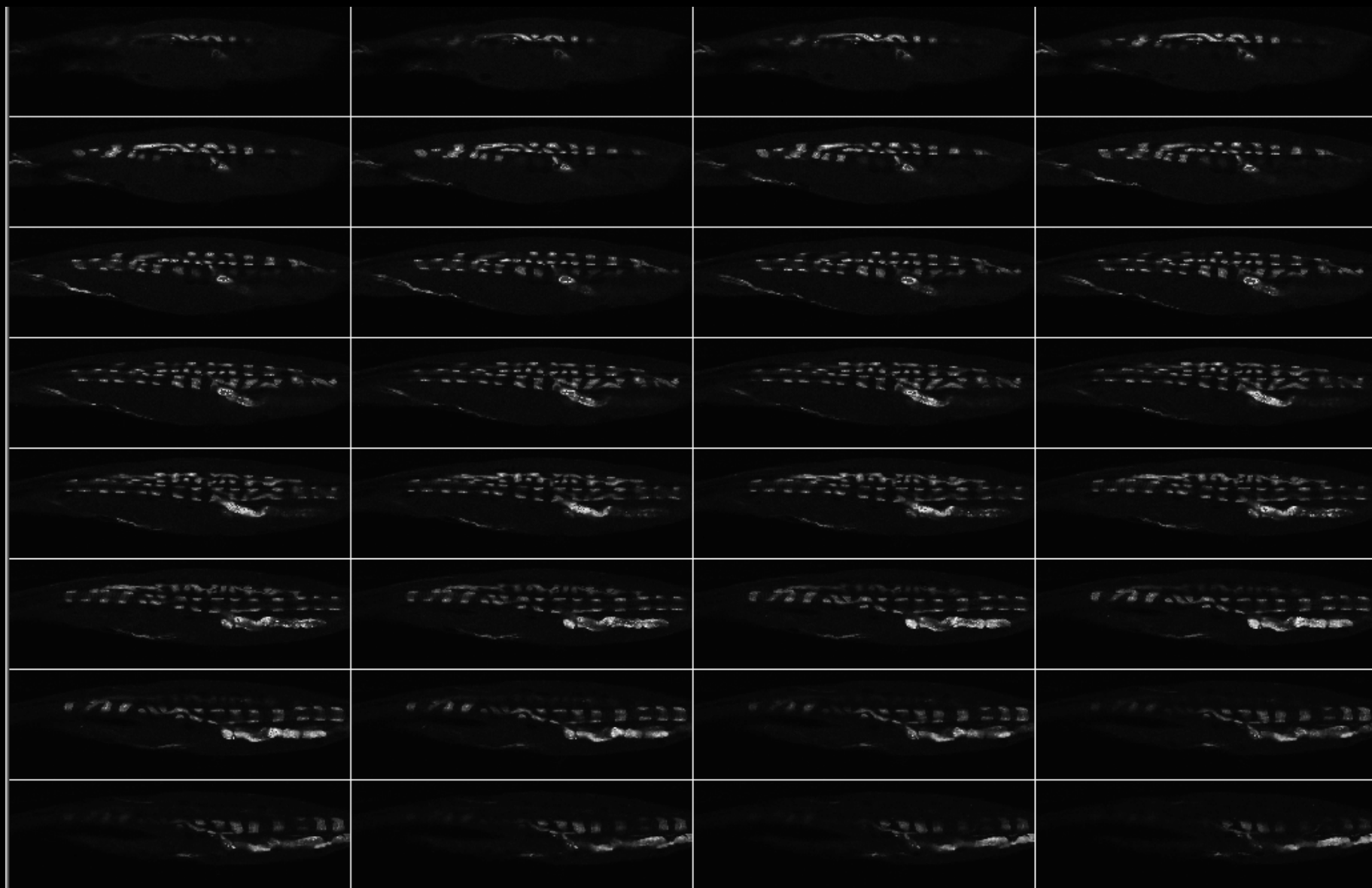
40x 1.3 NA 230 um



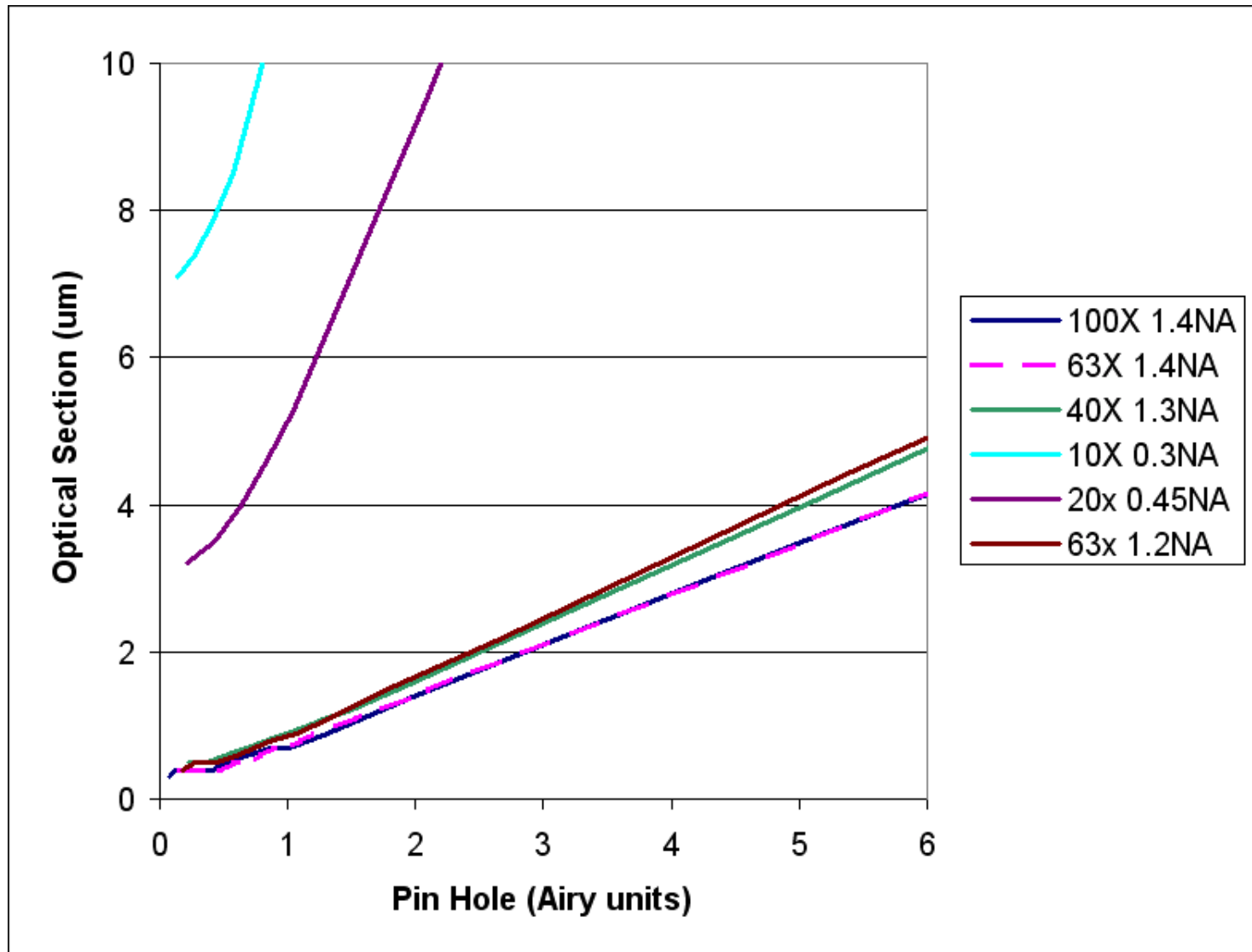
wide field

# Visualization - Focus to Different Planes

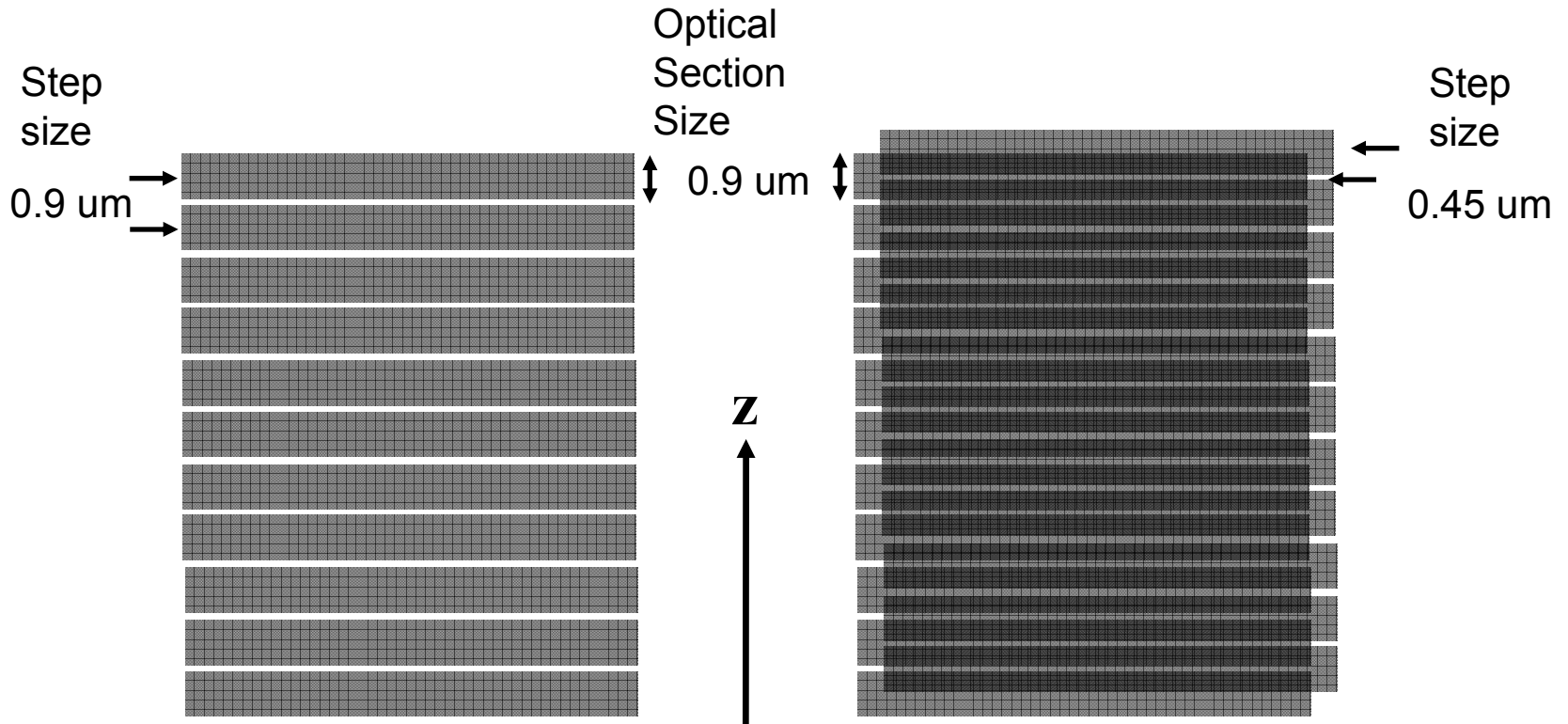
## Serial Sections



# Pinhole Size versus optical Section



# Optical Sectioning

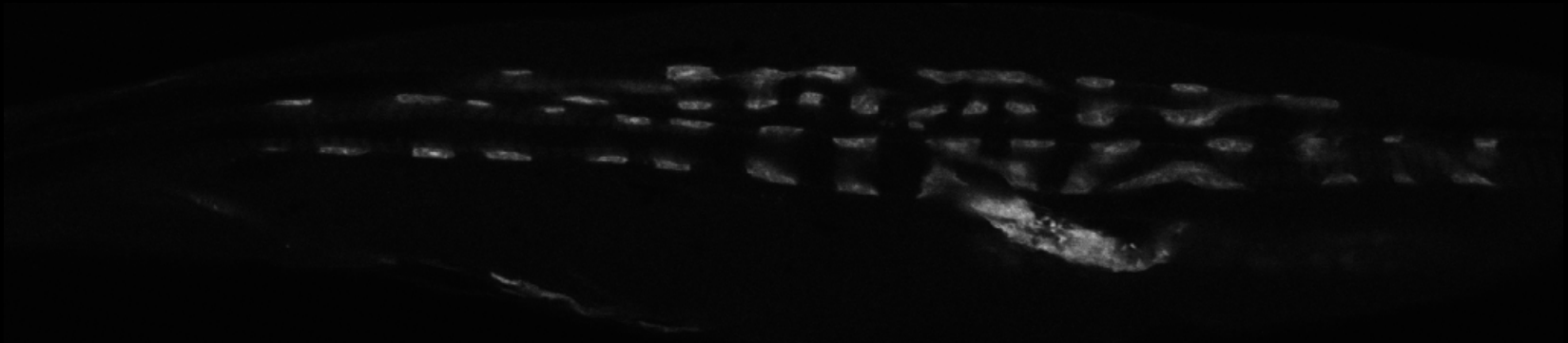


Inadequately  
Sampled

Just Adequately  
Sampled  
(Nyquist criterion at least twice  
frequency,  $\sim 2.3x$ )



# Visualization - Live Muscle Spindle (Thy1-YFP)

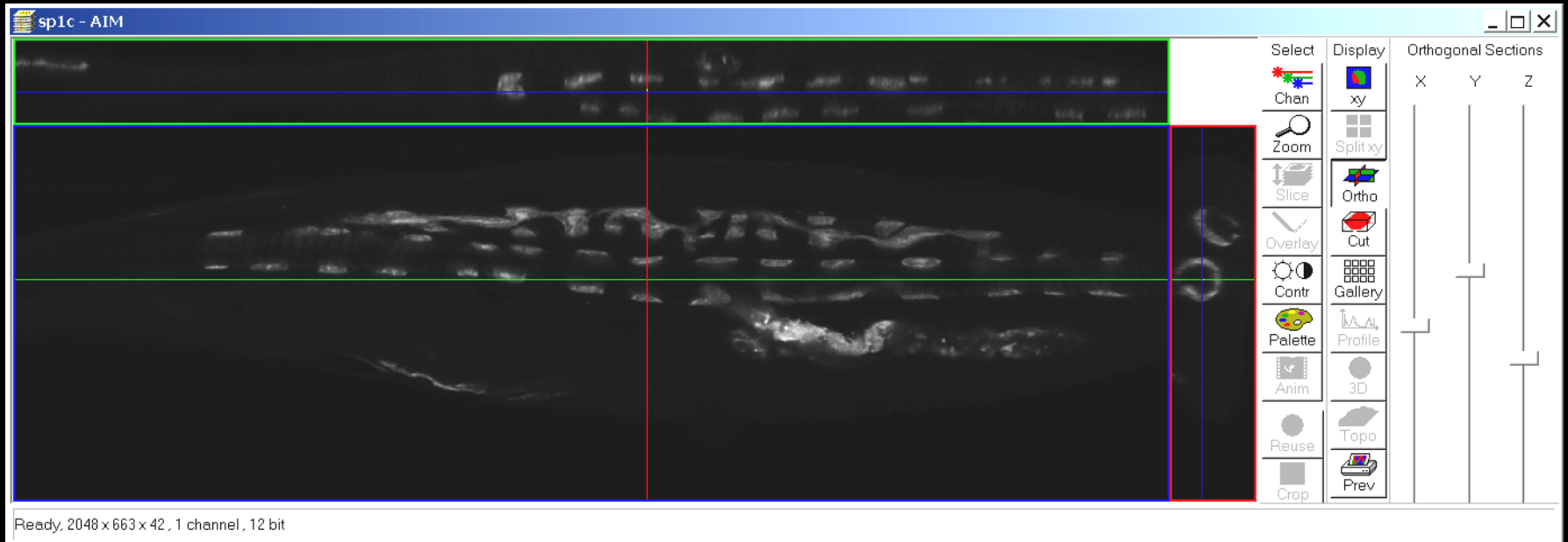


single slice

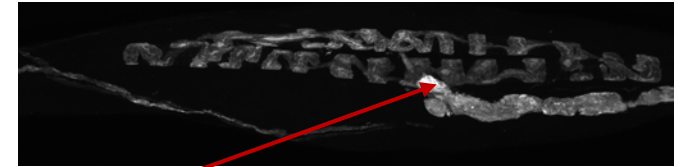
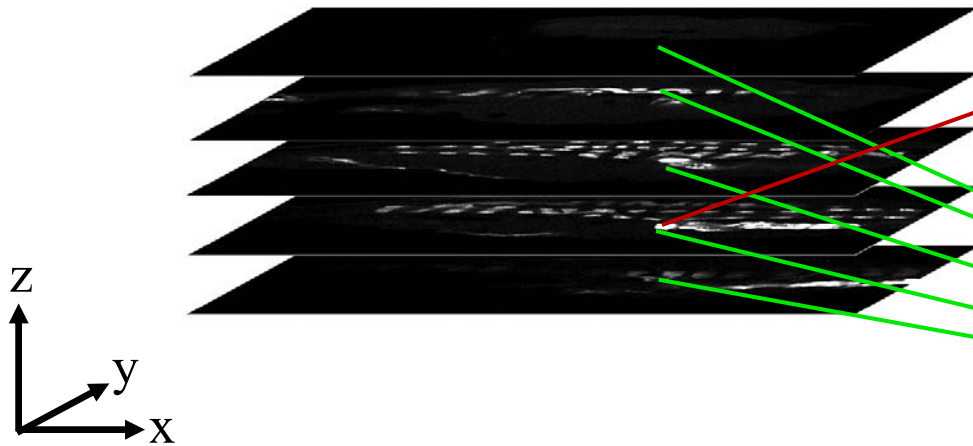
serial slices

40x 1.3 NA 230 um

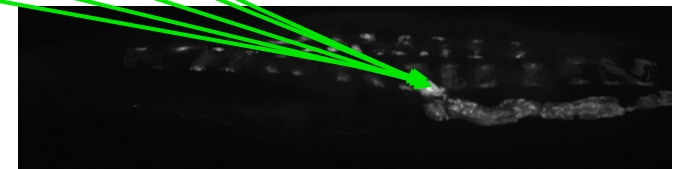
# Visualization - Orthogonal View



# Visualization - Extended Focus 40x

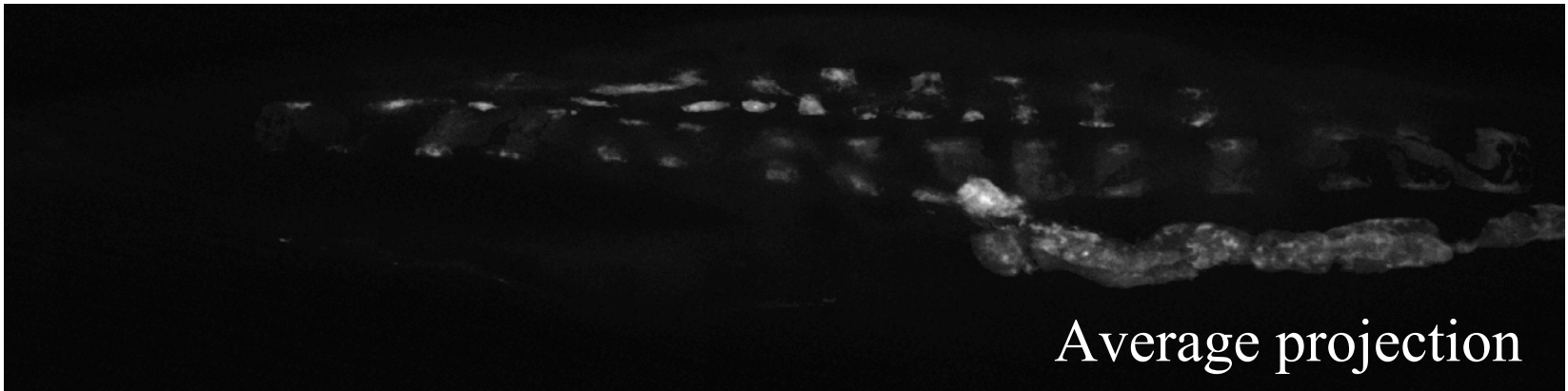
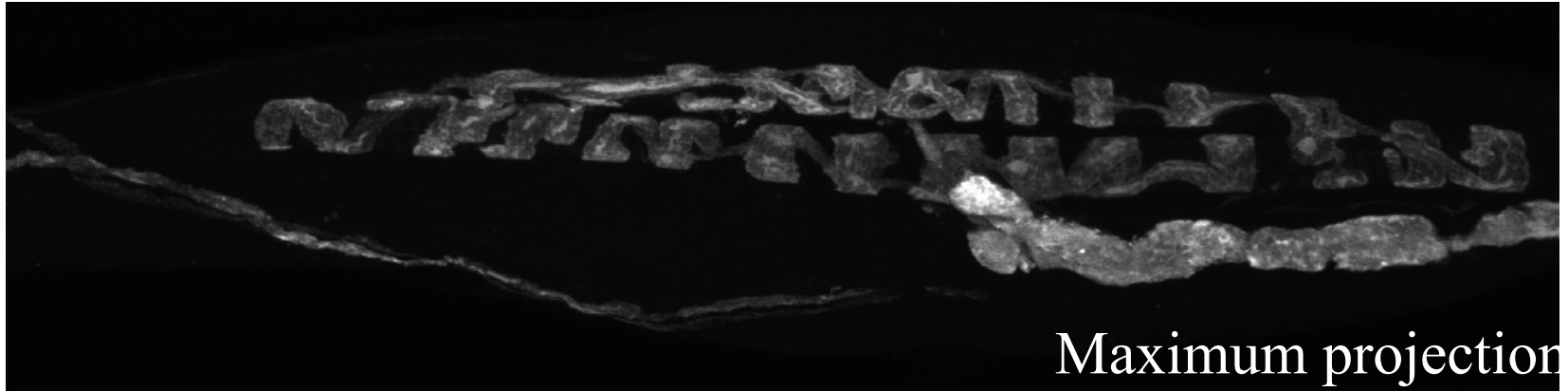


**Maximum projection**  
Brightest pixel in column of stack – good for viewing thin structures – bad for quantification



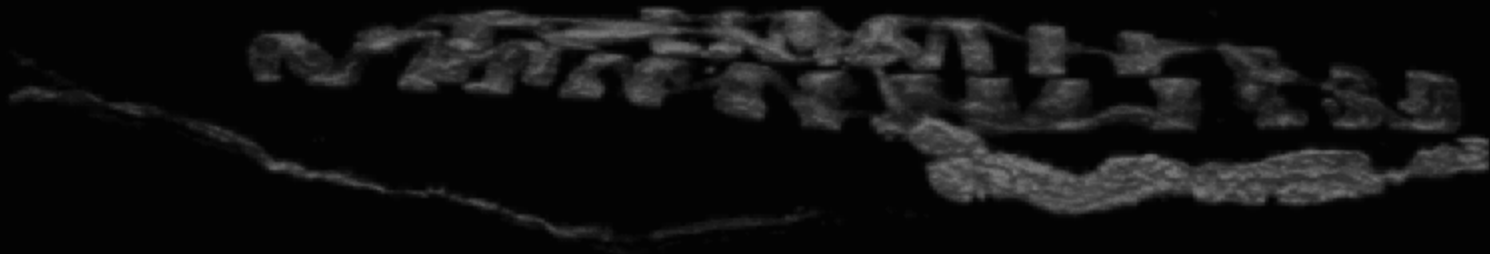
**Average projection**  
Sum corresponding pixels and divide by the number of slices – good for quantification

# Visualization - Extended Focus 40x

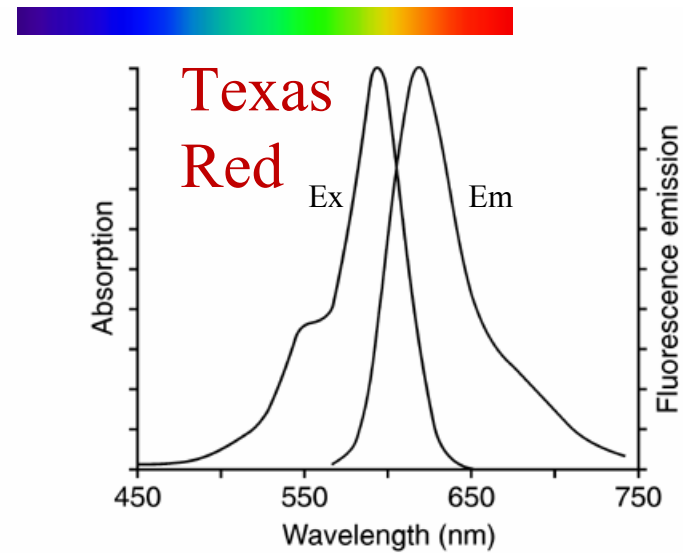
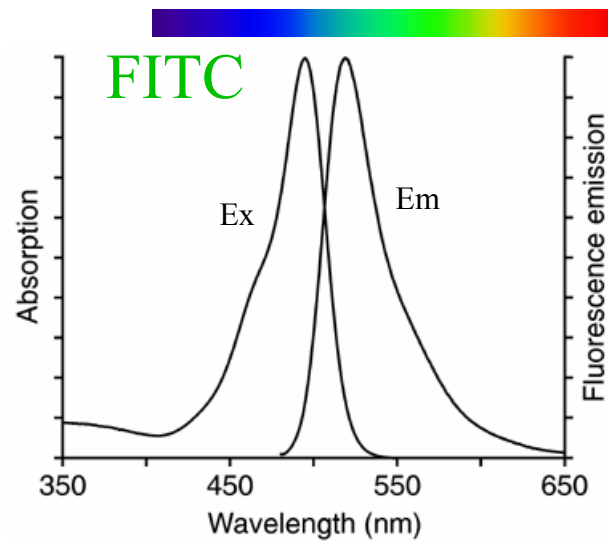


# Visualization - Volume Rendering

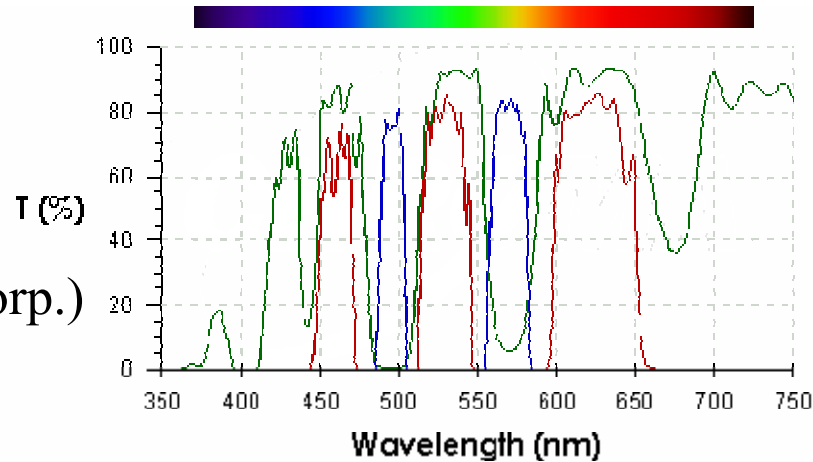
Each voxel – intensity \* opacity



# Confocal Multichannel Fluorescence

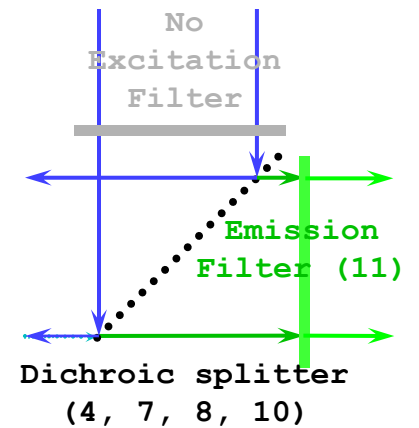
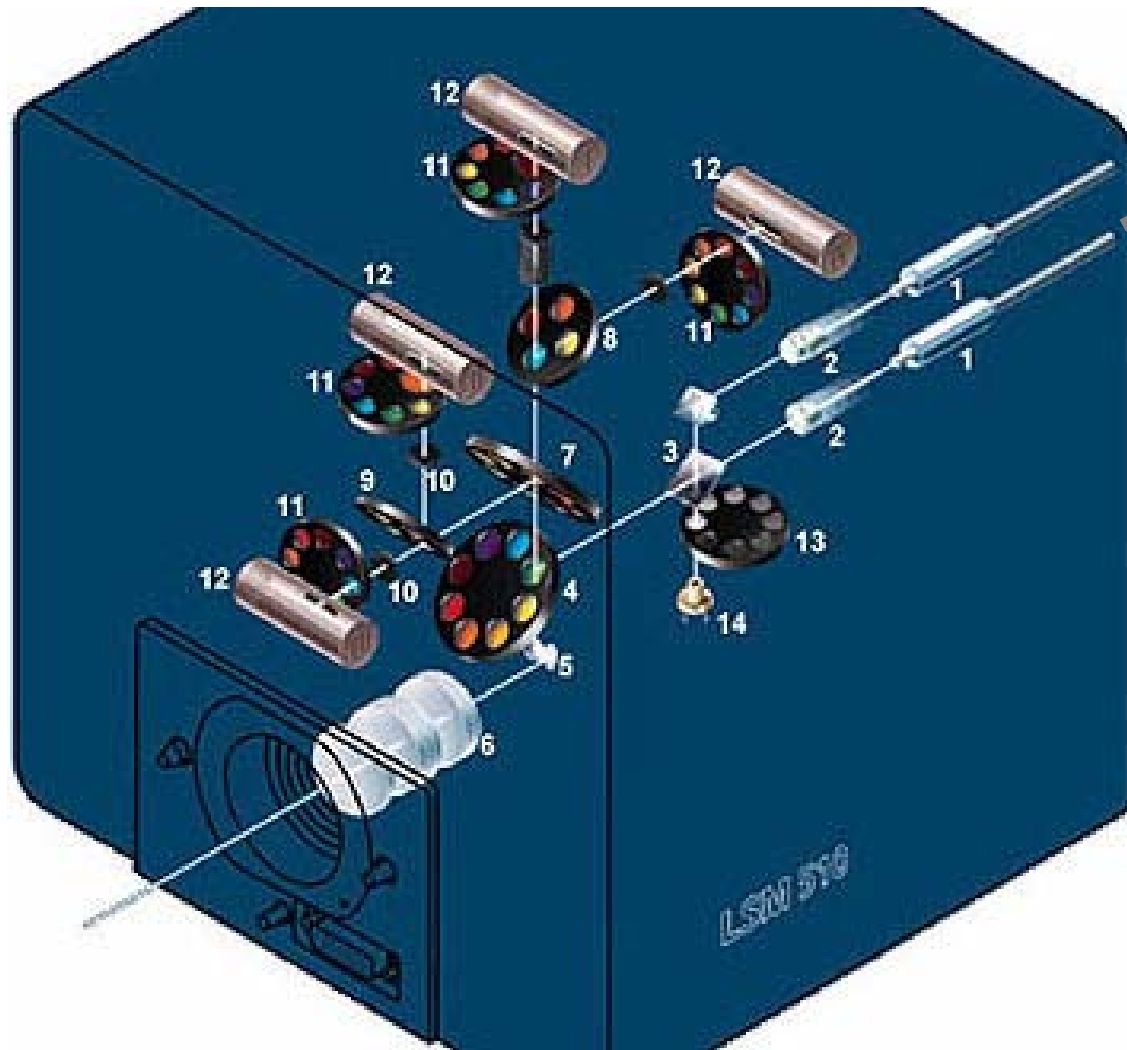


Triple band  
Filter (Chroma Corp.)

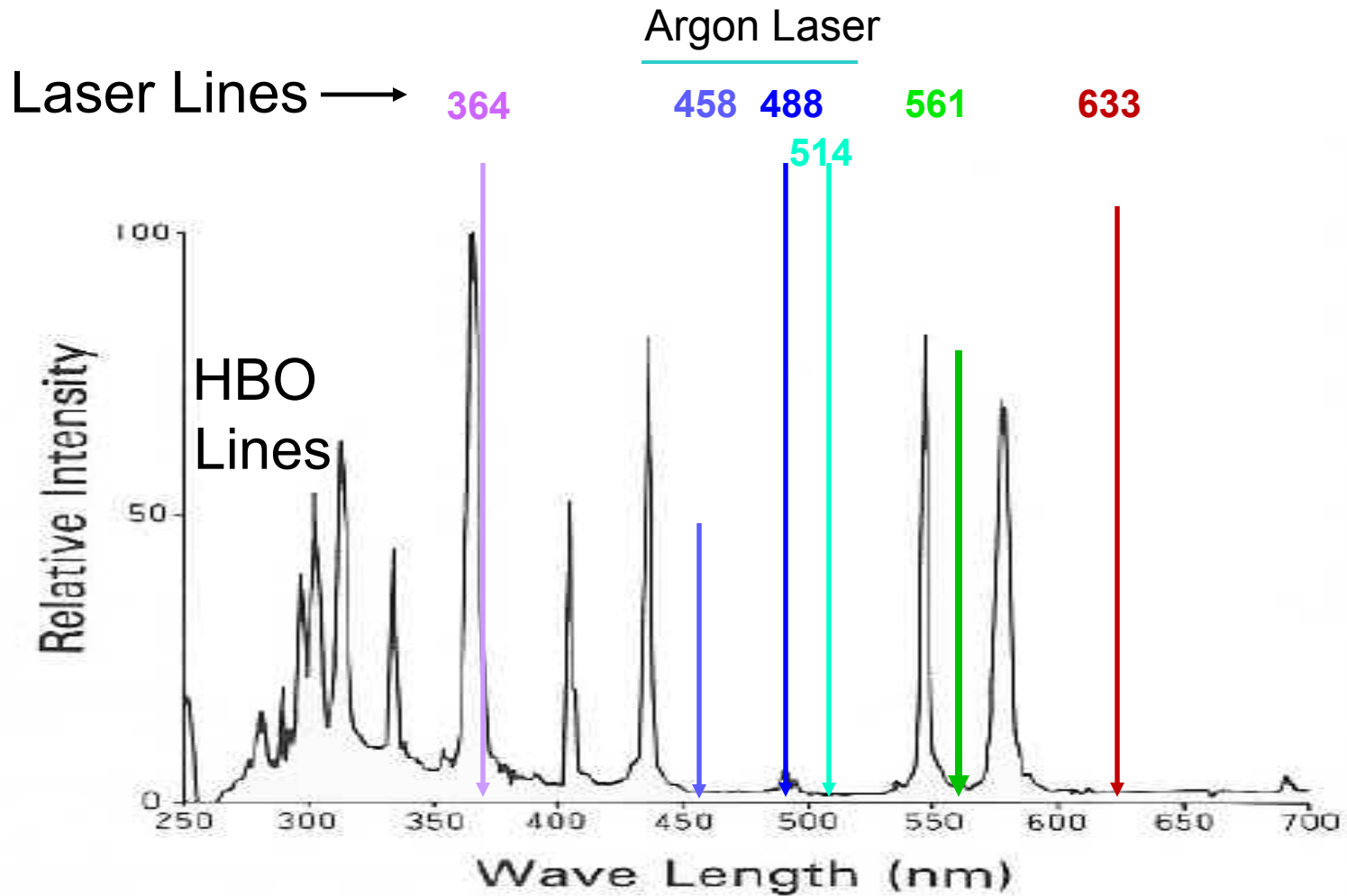


- Exciter
- Dichroic
- Emission

# Confocal Multichannel Fluorescence

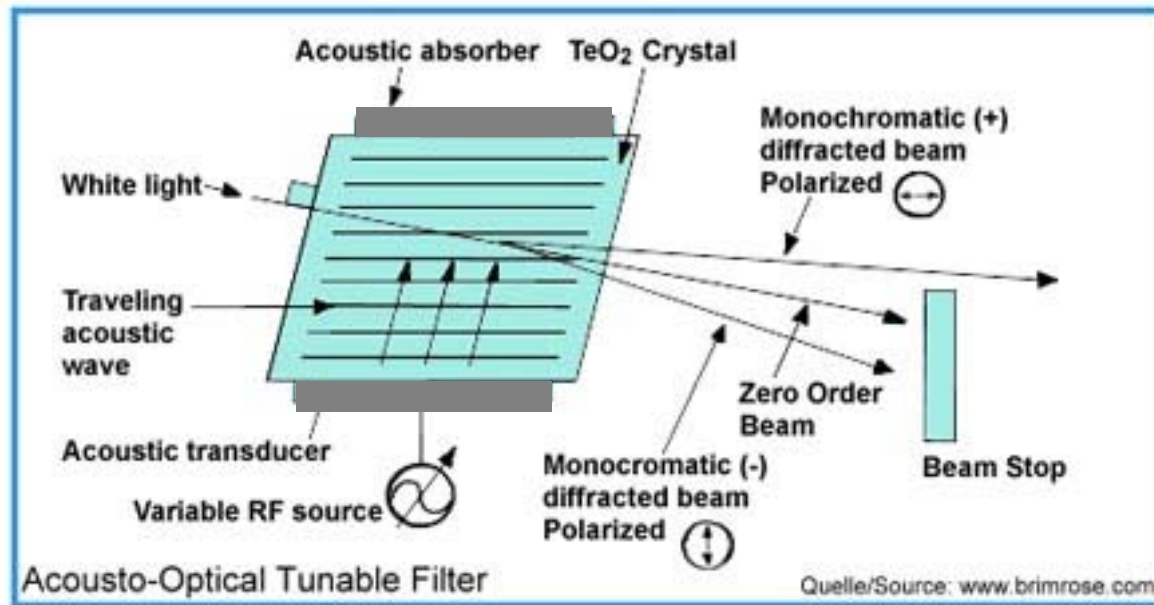


# ARC Lamp versus Laser Excitation



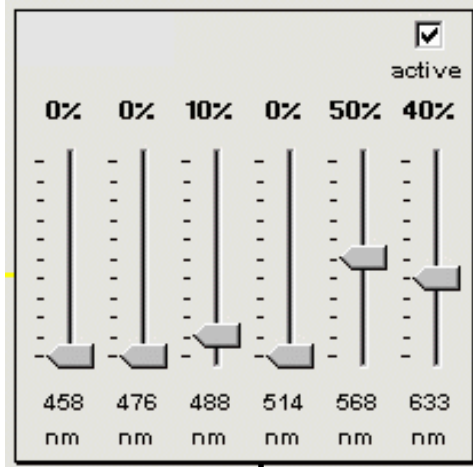


# Acousto-Optical Tunable Filter (AOTF)

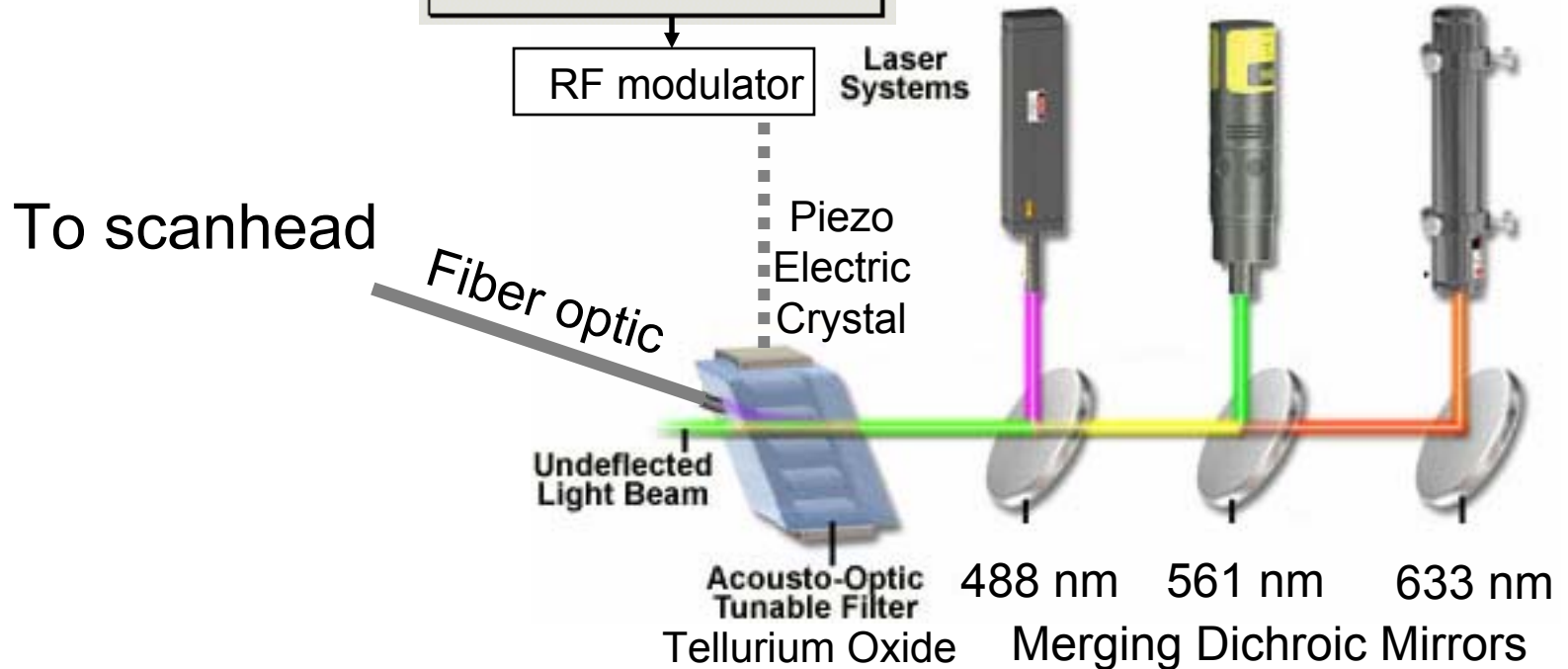


RF = radio frequency

# Excitation Light System – AOTF Controlled

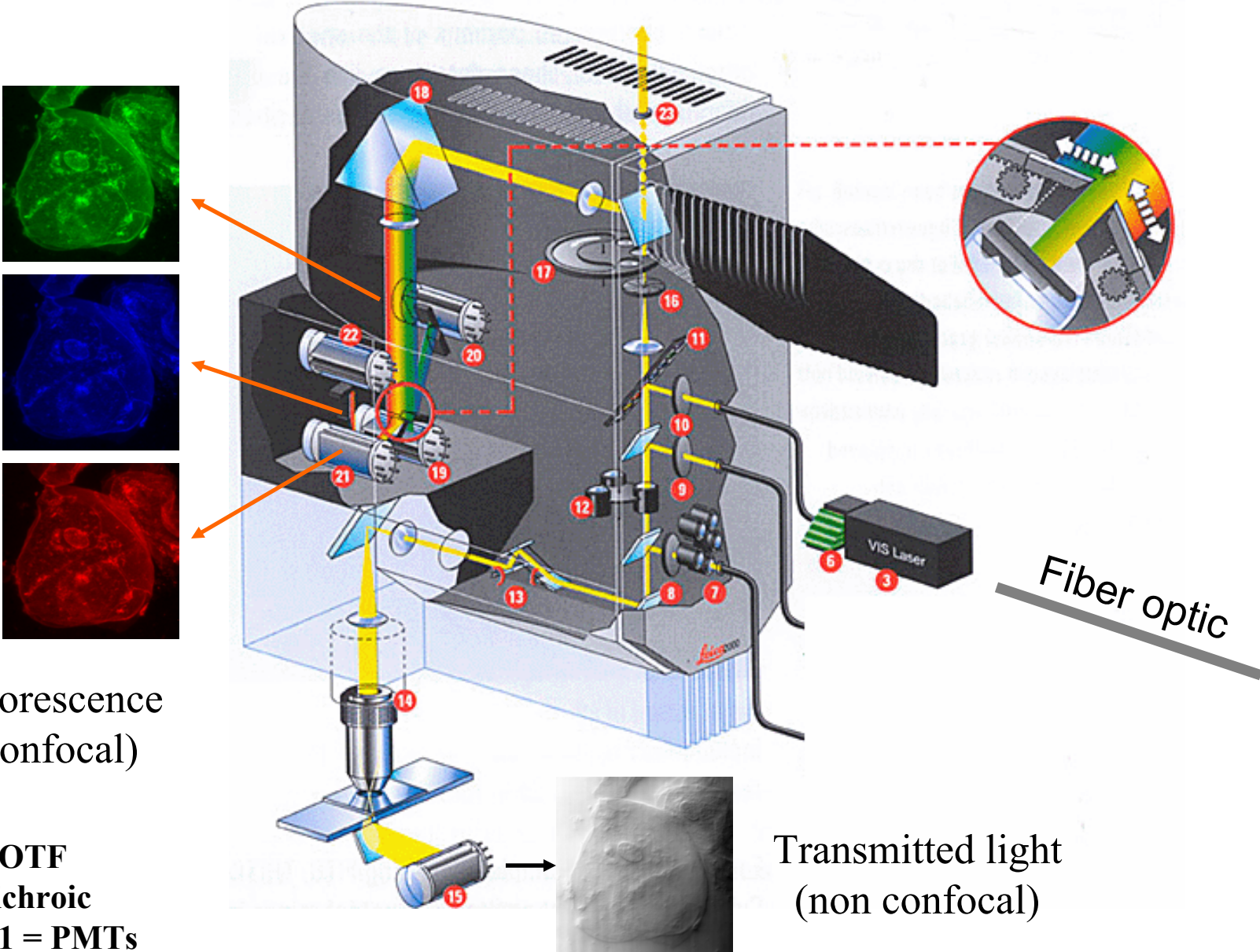


Replaces excitation filters



RF = radio frequency

# Confocal Multichannel Fluorescence

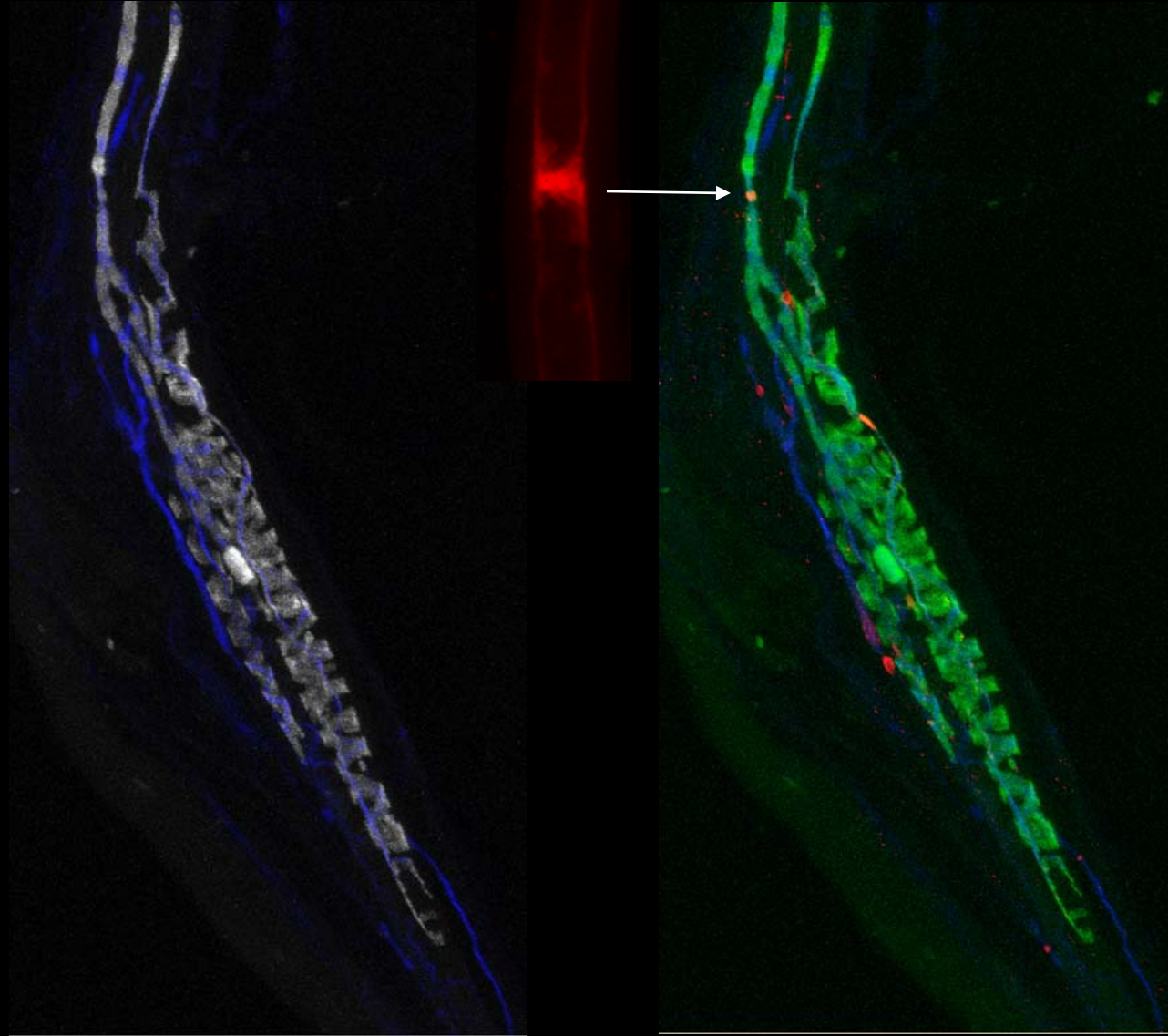


# Confocal Multichannel Fluorescence

Ab-Neurofilaments 200 (blue)

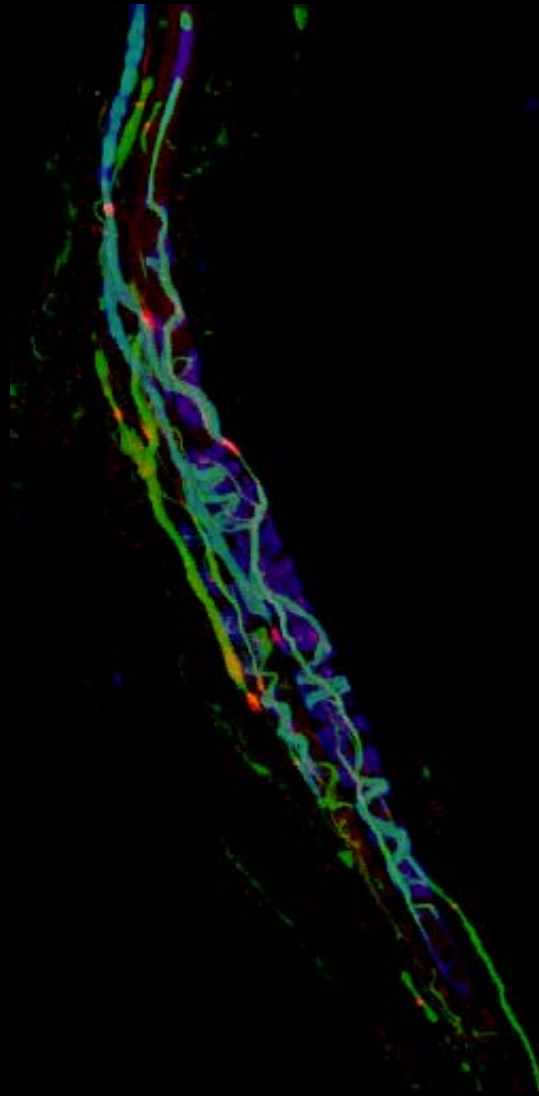
YFP (gray/green)

VNaCh (red)



# Confocal - Multichannel Fluorescence

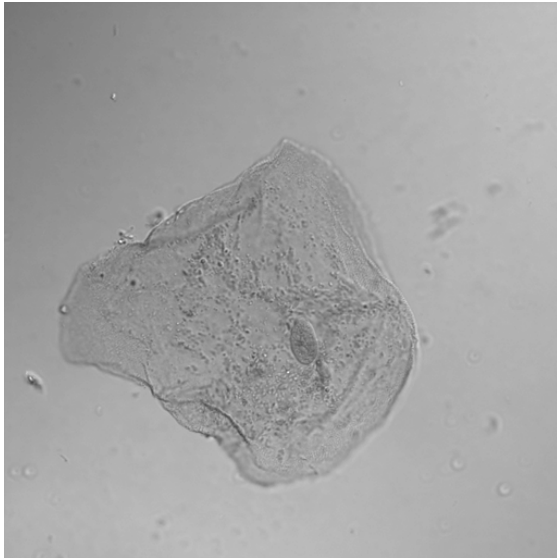
Ab-Neurofilaments 200 (green)    YFP (Blue)    VNaCh (red)



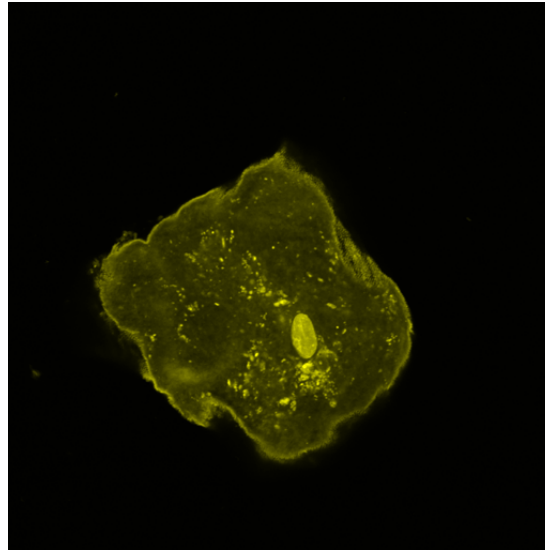
# Confocal Microscopy – Summary



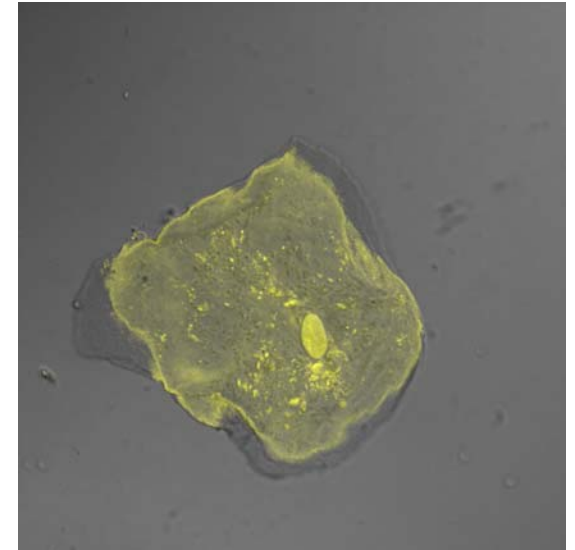
- A microscope system which removes out of focus information optically.
- Better lateral and much improved axial resolution
- Higher contrast
- Reduced glare
- Single voxel scanning permits 3-D reconstructions



(DIC) Transmitted  
(widefield)



Fluorescence  
FM 1-43 membrane dye  
(confocal)



Overlay

# References

- Confocal Microscopy Methods and Protocols, in Methods in Molecular Biology, Stephen W. Paddock, 1999
- Handbook of Biological Confocal Microscopy, 2<sup>nd</sup> ed., James Pawley, 1995
- Handbook of Biological Confocal Microscopy, 3<sup>rd</sup> ed., James Pawley, 2006
- Confocal Microscopy for Biologists, Alan R. Hibbs, 2004  
(Missing at UNC, Duke & NC State)

# Advanced Fluorescence/Confocal

## May 14

- Multi-channel bleed through/cross talk
- Co-localization
- Live cell imaging
- FRAP, fluorescence recovery after photobleaching
- Deconvolution
- **Your requests?**

